

43 barbers and their clients are constantly being exposed to bacterial or fungal contamination during
44 their services (Tharmila *et al.*, 2012). Naturally human hair harbours many pathogenic bacteria,
45 and also it acts as a potential source of cross infections. Bacteria such as *Staphylococcus aureus*,
46 *Escherichia coli*, *Streptococcus viridians*, β haemolytic streptococci, the *Proteus* group,
47 *Pseudomonas pyocyanea* and *Streptococcus faecalis* have been reported to be present in the hair
48 (Tharmila *et al.*, 2012). This therefore represent a potential risk factor for customers and visitors
49 to the salon. Due to these problems, in this study, we aimed to investigate the antibiotic
50 **susceptibility** pattern of bacterial pathogens isolated from hair in barbing salons, within Benin
51 metropolis.

52 **MATERIALS AND METHODS**

53 **Samples collection**

54 Hair samples were randomly collected from ten (10) different barbing salons within Benin
55 metropolis, Edo State, Nigeria. **The ten salons were carefully selected based on population of**
56 **clients that always go there. Before sampling was carried out, appropriate arrangement was**
57 **ensured to disinfect the perimeter inside the barber's chop in order to prevent contamination.** The
58 samples were collected with sterile spatula and placed in a sterile universal container to avoid
59 contamination. Samples were then transported to the laboratory for microbial analysis without
60 delay.

61 **Culture medium and Isolation of Bacteria**

62 Commercially available Nutrient agar medium was obtained and prepared following the
63 manufacturer's instructions. Ten gram (10g) of each sample was weighed and aseptically
64 introduced into 90ml of sterile distilled water, properly shaken before a 10 fold serial dilution, up
65 to 10^{-3} , was performed. Pour plate isolation method was used for microbial enumeration. In this
66 method, 0.1ml from each dilution was pipetted into sterile Petri dish and labelled. About 20ml
67 of prepared agar medium was dispensed into the various Petri plates and mixed. The nutrient
68 agar plates were allowed to solidify and then incubated at 37°C for 24 hours, after which the
69 developed colonies were counted to obtain total viable count. Discrete distinct colonies were
70 purified by subculturing into nutrient agar plates using the streak plate method.

71 **Procedure for identification of the organisms**

72 The bacterial isolates were characterized and identified based on their cultural characteristics and
73 biochemical reaction as presented in table 2.

74 **Antibiotic Susceptibility pattern of the isolates:**

75 Antimicrobial disc tests were performed on the isolates using the following antibiotic discs:
76 perfloracin, gentamicin, ampiclox, zinnacef, amoxicillin, rocephin, ciprofloxacin, streptomycin,
77 erythromycin, gentamycin, septrin, chloramphenicol, sparfloracin, and ofloxacin. The organism
78 was inoculated into nutrient broth in test tube and incubated for 24hours. Measured 0.1 ml of
79 liquid culture was added to solidified nutrient agar in Petri dish and a glass spreader was used to
80 even spreading on the agar surface. The plates were allowed to dry for 5-10 minutes, after which
81 standard antibiotics disks was layered on the inoculated agar. The plates were incubated at 37°C
82 for 24hours. Clear zones around each disk **were** measured and interpreted as either resistance or
83 **sensitive**.

84 **RESULTS**

85 Table 1 show total bacterial counts of the different hair samples. Value ranged from
 86 $2.80 \times 10^3 \pm 0.8 \text{ cfu/g}$ to $6.13 \times 10^3 \pm 0.21 \text{ cfu/g}$. Table two describes the cultural, morphological and
 87 biochemical characterization of the bacterial isolated. The isolates identified include *Escherichia*
 88 *coli*, *Proteus vulgaris*, *Streptococcus viridians* and *Corynebacterium* sp. Figure 1 presents
 89 percentage distribution of the bacteria species among the different samples with the most
 90 prevalent being *Corynebacterium* sp (80%) while the least was *Escherichia coli* (40%). Table 3
 91 explains the antibiotic sensitivity pattern of bacterial isolates. All identified bacterial strain were
 92 observed to be multiple drug resistant.

93
 94 Table 1: Total viable bacterial counts in hair samples from dilution of 10^{-2}

Samples	mean± SE(x10 ³ cfu/g)	P- value
A	2.80±0.8 ^a	0.000
B	5.77±0.31 ^b	
C	5.80±0.27 ^b	
D	6.07±0.21 ^b	
E	5.03±0.15 ^c	
F	6.13±0.21 ^b	
G	5.10±0.20 ^c	
H	5.23±0.25 ^c	
I	5.53±0.68 ^b	
J	4.27±0.21 ^d	

95 Key: A-J = Hair from barbing salon in ten different locations in Benin City

96 SE = Standard error; P<0.05

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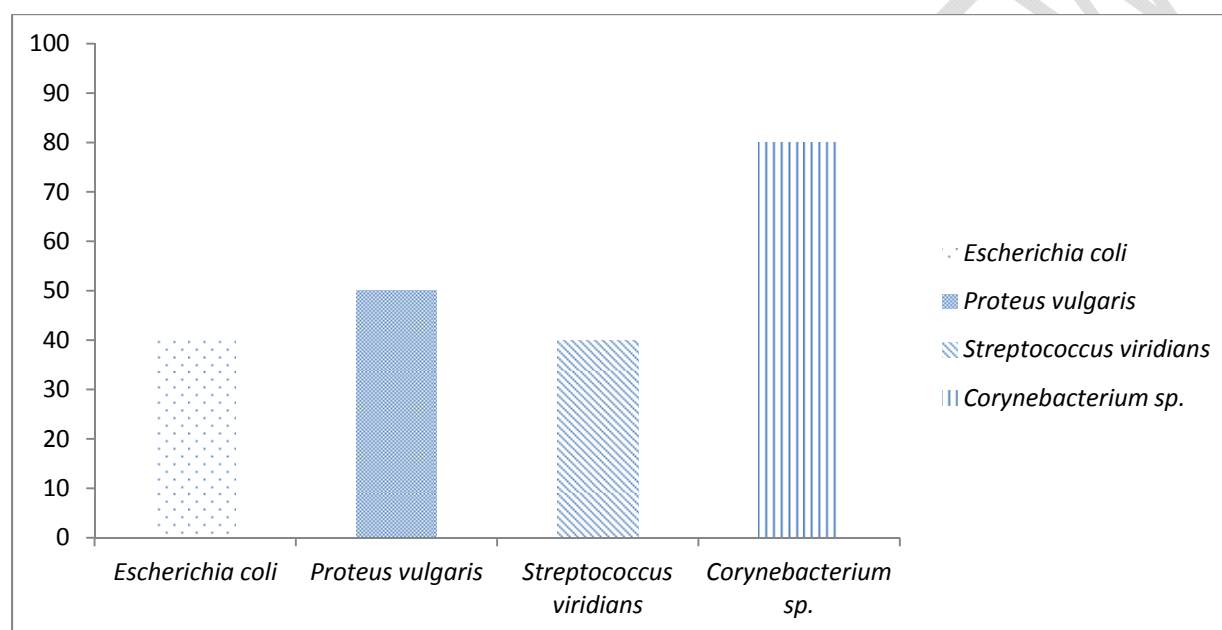
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99 **Table 2: Cultural, morphological and biochemical characteristics of the bacterial isolates**

Characteristics	Isolates			
	B1	B2	B3	B4
Cultural				
Elevation	Low convex	Flat	Convex	Convex
Margin	Entire	Undulated	Entire	Entire
Colour	Cream	Cream	White	Cream
Shape	Circular	Irregular	Circular	Circular
Size	Small	Medium	Small	Medium
Morphological				
Gram staining	-	-	+	+
Cell type	Rod	Rod	Cocci	Rod
Cell arrangement	Single	Single	Chains	Single
Biochemical				
Catalase	+	+	-	+
Oxidase	-	-	-	-

Coagulase	-	-	-	-
Urease	-	+	-	+
Indole	+	+	-	-
Citrate	-	+	+	+
Sugar fermentation				
Glucose	+	+	+	+
Lactose	+	-	-	-
Possible isolates	<i>Escherichia coli</i>	<i>Proteus vulgaris</i>	<i>Streptococcus viridians</i>	<i>Corynebacterium sp.</i>

100



101

102 Fig. 1: Prevalence of bacterial isolated from hair

103 Table 3: Antibiotic susceptibility pattern of isolated bacteria

Bacteria	No. I	Antibiotics									
		CPX	St	SXT	E	PEF	CN	APX	Z	AM	Ro
Gram +ve											
<i>S. viridians</i>	4	2(50)	2(50)	3(75)	1(25)	4(100)	4(100)	3(75)	2(50)	3(75)	4(100)
<i>Corynebacterim sp.</i>	8	3(37.5)	4(50)	6(75)	3(37.5)	2(25)	2(25.0)	5(62.5)	1(12.5)	4(50.0)	3(37.5)
Gram -ve											
<i>Escherichia coli</i>	4	4(100)	1(25)	0(0.0)	0(0.0)	0(0.0)	2(50)	1(25)	3(75)	1(25)	0(0.0)
<i>Proteus vulgaris</i>	5	0(0.0)	0(0.0)	3(60)	2(40)	4(80)	1(20)	1(20)	3(60)	1(20)	1(20)

104 **KEY:**

105 No. I= Number of isoaltes; CPX-Ciprofloxacin, Ro-Rocephin, St-Streptomycin, AU-Augmentin,
106 SXT-Septrin, SP- Sparfloxacin, E-Erythromycin, CH-Chloramphenicol, PEF-Pefloxacin, CPX-
107 ciprofloxacin, CN-Gentamicin, APX-Ampiclox, AM-Amoxicillin, Z-Zinnacef

108

109 Discussion

110 High bacteria load were observed in the different hair samples from different barbing salons.
111 Total bacterial counts ranged from $2.80 \times 10^3 \pm 0.8$ cfu/g to $6.13 \times 10^3 \pm 0.21$ cfu/g. Ajuzie and
112 Osaghae (2011) reported high bacterial counts from salon waste water. The bacteria may have
113 come from washed hair. Variations in bacterial counts from the different samples reflects the life
114 style of the individual and the kind of hair treatment. These high bacteria counts shows that
115 human hair is highly contaminated with diverse microorganisms especially bacterial, some of
116 which can be potential pathogens of public health importance (Yun *et al.*, 2010). This finding
117 means that human hair in barbing salon represent potential source of bacterial contamination of
118 either food or water. Also due to the light nature of the hair, it can be easily blown by wind to
119 surrounding environment where it may deposit on food or water system, thereby leading to
120 contamination.

121 Based on the cultural, morphological and biochemical characterization of the isolates, four
122 different bacterial species were isolated and they included *Escherichia coli*, *Proteus vulgaris*,
123 *Streptococcus viridians* and *Corynebacterium* sp. Enemuor *et al.* (2013) reported on the
124 prevalence of these bacterial strains in hair dressing and beauty salons. Summers (1995) stated in
125 his work that hair is a reservoir of *Staphylococcus aureus*. Although *S. aureus* was not detected
126 in this work, the isolated bacterial strains from this work are potential pathogens implicated in
127 various diseases of humans. *E. coli* is known to cause various gastrointestinal disorder such as
128 diarrhoea; urinary tract infections and meningitis. *Proteus* spp. have been implicated in urinary
129 tract infections. *Streptococcus* spp. are causative agents of several human diseases including
130 pneumonia, caries and other pyogenic infections. The organism also produce super antigen
131 which hyper regulate T-cell proliferation and activation, leading to autoimmune diseases.
132 *Corynebacterium* sp. is a known human pathogen, causing diseases such as diphtheria. These
133 pathogens can easily be transmitted from one person to another most especially when one clipper
134 or comb is used for multiple customers. This calls for awareness on the part of customers, on the
135 possibility of being infected. Tharmila *et al.* (2012) investigated the inhibitory effect of some
136 traditional hair washing substances on hair borne bacteria, thus confirming the presence of
137 bacterial pathogens on human hair. In another research study, five bacterial isolates including
138 *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Streptococcus* sp, *Enterococcus* species and
139 *Enterobacteria* were reported (Enemuor *et al.*, 2013). The presence of these potential pathogens
140 is an indication that hairdressing and beauty salons could be contributing to the spread of
141 infection within the community (Enemuor *et al.*, 2013). Infection can occur during hairdressing
142 procedures since items such as razors, scissors, combs, clippers and hairpins can accidentally penetrate
143 the skin. Blood and body fluids do not have to be visible on instruments, equipment or working surfaces
144 for infection to be transmitted. Bacterial Infections that can be spread in hairdressing premises include
145 skin infections on the scalp, face and neck such as impetigo (Brown, 2006; Amodio *et al.*, 2010; Barn
146 and Chen, 2011).

147 Summers *et al.* (1995) reported the presence of *Escherichia coli*, *Streptococcus viridans*,
148 *Proteus* group, *Haemolytic streptococci*, *Pseudomonas pyocyanea*, *Streptococcus faecalis* and
149 *Staphylococcus aureus* from the hair of the scalp.

150 The different bacterial strains from this study were observed to be variedly distributed among the
151 different hair samples. The least occurring bacterial species were *Escherichia coli* and *Proteus*
152 *vulgaris* with percentage distribution of 40% each while the most widely distributed was
153 *Corynebacterium* sp. (80%).

154 Antibiotic susceptibility of the bacterial isolates revealed varying degree of resistance to
155 conventionally used antibiotics. All the isolates were observed to be multiple drug resistant.
156 Result revealed that isolated bacterial from hair, were resistant to multiple antibiotics. There
157 were variations in their degree of antibiotic resistance. Of the four isolates of *Streptococcus*
158 *viridans*, 2(50%) were resistant to ciprofloxacin, streptomycin, zinnacef, 3(75%) were resistant
159 to septrin, ampiclox and amoxicillin, while 4(100%) were resistant to perfloxain, gentamicin
160 and rocephin. Antibiotic resistant pattern of *Corynebacterium* sp revealed that 5(62.5%) were
161 resistant to ampiclox while 6(75%) were resistant to septrin. Perfloxacin, gentamicin and
162 zinnacef were highly effective against *Corynebacterium* sp in this study. The 4(100%) of
163 *Escherichia coli* were sensitive to augmentin, ofloxacin, septrin and ciprofloxacin. However,
164 they were resistant to chloramphenicol, perfloxacin and streptomycin. *Proteus vulgaris* was also
165 sensitive to chloramphenicol, sparfloxacin and resistant to augmentin, septrin and streptomycin

166 Antibiotic resistant genes in bacterial have been shown to be borne on either plasmid or
167 chromosomally mediated. Bacterial pathogens have been reported to use various mechanisms to
168 resist antibiotics, such mechanisms include use of efflux pumps, drug inactivating enzymes, drug
169 modifying enzymes among others.

170 **Conclusion**

171 Hair samples from barbing salons have been shown to be highly contaminated with bacterial
172 isolates. The isolated bacteria were found to be bacterial pathogens that are implicated in many
173 human and animal diseases. These pathogens were also observed to be multidrug resistant. It is
174 highly recommended that individual that goes to barbing salons should have their own clipper
175 and always disinfect it to reduce the microbial load. People should also be aware of the potential
176 possibility of pathogen transmission in barbing salon especially when such salon is situated near
177 water or food canteens.

178

179 **Competing Interests**

180 All authors have declared that no competing interests exist.

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