

An Over View of Dermatophytosis in Rabbits

Abstract:

Dermatophytosis is a fungal infection of the skin caused by dermatophytes-filamentous fungi which have ability to invade the epidermis and keratinized structure derived from it such as hair or nails. Rabbits are one of dermatophytes host; young rabbit below 12 months of age were more frequently affected with the disease. *T. mentagrophytes* is the most common dermatophytes isolated species. The disease can be diagnosed by direct examination, fungal culture, skin biopsy sero and molecular diagnosis methods. This overview forecast more light of the different aspects of this disease.

Key words:

Dermatophytosis,Rabbit,Clinical feature, Diagnosis.Treatment

Introduction:

Rabbits are calm by nature. They are prone to many bacterial, fungal or parasitic skin diseases if proper care is not taken. Among them dermatophytosis is one of the most common diseases [1]. Dermatophytosis is a superficial cutaneous infection with one or more of the fungal species in the keratinophilic genera *Microsporum*, *Trichophyton*, or *Epidermophyton* [2, 3]. Young or immune compromised rabbits are most susceptible to the disease [4]. Dermatophytosis is a zoonotic disease so it has important implications in public health [5]. Infection with dermatophytosis can be occurred in receptive hosts via arthrospores present on the hair coats of infected animals or in the environment [6].

Epidemiology:

The possibility of infection of dermatophytosis depends on fungal species, host age, immunocompetence, condition of exposed skin surfaces, host grooming behavior, and nutritional status [6,7]

Young below 12 months of age or immune compromised rabbits are thought to be most susceptible [8]. However, differences in skin secretions, especially lower levels of fungistatic

31 fatty acids in sebum and lower levels of fungal inhibitory sphingosine, and the fast growth and
32 replacement of hair may also play a role in facilitating infection [9]. The presence of ecto-
33 parasites, especially fleas and *Cheyletiella* mites, can also lead to spread of dermatophytosis [9].

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35 **Risk factors:**

36 1- Young animal

37 2- Overcrowding

38 3- high humidity

39 4- poor sanitation

40 5- malnutrition

41 6- Immunosuppression (including immunosuppressive treatment)

42 7- Injury by ectoparasites or scratches due to pruritus

43 Reported by [10]

44 **Transmission**

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46 Dermatophytosis can be transmitted by direct or indirect contact with infected hair, scales or
47 materials. Infectious microconidia in the environment or on fomites can persist for many months.

48 The pathogenesis of dermatophytosis includes several stages; adhesion, germination, invasion,
49 penetration. Natural defences against dermatophytes depend on both immunological and non-
50 immunological mechanisms so infectious microconidia must first overcome a couple of local
51 defenses to be able to adhere the keratinized tissue, the stratum corneum [11,12].

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53 [11].

54 **Clinical features:**

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57 Clinically, dermatophytes infect the epidermis and adhering structures, including hair follicles
58 and shafts [13, 14]. In rabbits dermatophytic infections may cause alopecia, redness scaly and
59 scurf localized mainly on the face, head, auricles, and dorsal area of the neck [15-17]. This
60 disease can also result in rabbit malnutrition, growth retardation, feed remuneration reduction
61 and even death.

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67 **Fig.1: Alopecia in mouth. (17)**
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70 **Fig.2: Localized skin lesion of dermatophytes on leg (21)**

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Fig.3: Numerous canary crusts and large scales were present on the head of rabbits with hair loss.

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Etiology:

T. mentagrophyte is the most common dermatophytes isolated from rabbits and some researchers consider rabbits as asymptomatic carriers of this organism [18-22]. *M.gypseum* was also isolated from rabbits [23]. *M.canis* was reported by [24]. *T.verrucosum*. *Arthroderma benhamiae* were also recorded [25]. Rabbits are reported to be carrier for dermatophytes [26] so isolated *T. mentagrophyte*, *M.gypseum*, *M.nanum* and *M.canis* from healthy rabbits [27]

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Diagnosis:

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Hair and scrapings samples were collected with forceps or scalpel just behind the extending margin in the infected area. Samples can be kept in polyethylene bags. [28]

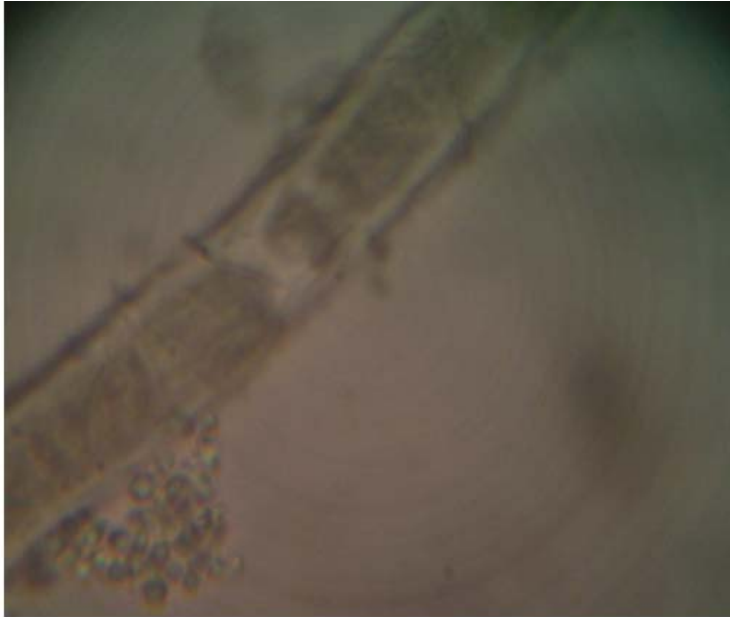
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Direct examination:

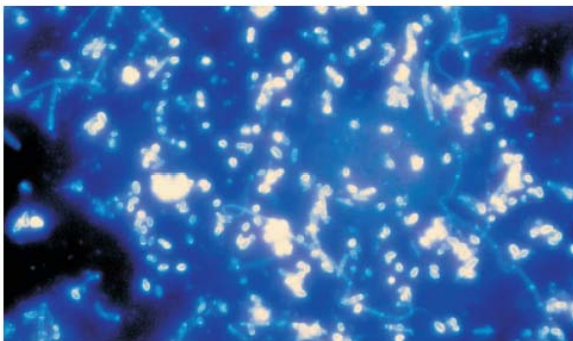
Hairs and scraping samples can be mounted in potassium hydroxide (KOH) of varying concentrations [29-31]. Infected hairs appear pale, wide and filamentous compared with normal

94 hairs when microscopically examined at x4 or x10 magnification. Arthrospores can be visible on
95 high magnification (x40). Positive result of KOH direct test can lead to positive cultures, which
96 are considered as the gold standard. Calcofluor white (a textile brightener) as an alternative to
97 KOH can be used because it binds specifically to the fungal cell wall and fluoresces strongly
98 when viewed under a fluorescence microscope [27].

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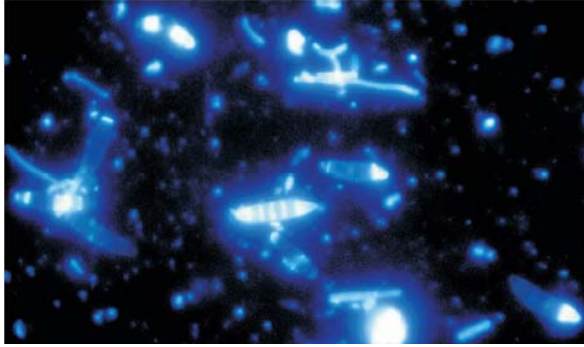


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102 **Fig. 4: Ectothrix arthrospores infection in hair**
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106 **Fig.5: Fluorescent microscopy (calcofluor white stain) of *Trichophyton mentagrophytes***
107 **complex hyphae and conidia isolated from healthy rabbits**
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113 **Fig. 6: Fluorescent microscopy (calcofluor white stain) of *Microsporium gypseum* hyphae**
114 **and macroconidia isolated from healthy rabbits**

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117 **Fungal culture:**

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119 Fungal culture is considered the ‘gold standard’ for diagnosis [32]. Sabouraud’s dextrose agar
120 (SDA) containing cycloheximide, penicillin and streptomycin were used in most diagnostic
121 laboratories. Plates should be incubated at 25°C for 5 weeks. Dermatophytes test media (DTM) is
122 recommended as the best media for isolation of dermatophytes because the presence of the red
123 color indicated positive result, this can help in early identification of highly suspected cultures
124 [33]. The isolates should be examined macroscopically and microscopically after staining with
125 lactophenol cotton blue using wet mount technique [34].

126 In addition to technique steps mentioned above, pigment production on corn meal agar, urease
127 activity on urea agar base, growth at 37°C on SDA.

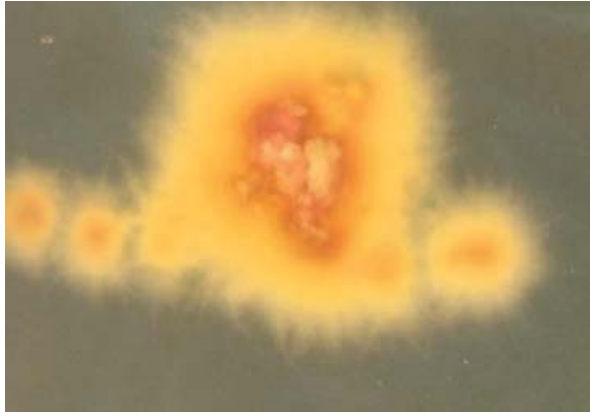
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130 **Fig.7: Culture of *T. mentagrophytes*: surface of colony show powder-like shape, white, loose**
131 **irregular mycelium on the edge.**

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Fig.8: Back side of *Trichophyton mentagrophytes*: pale yellow color



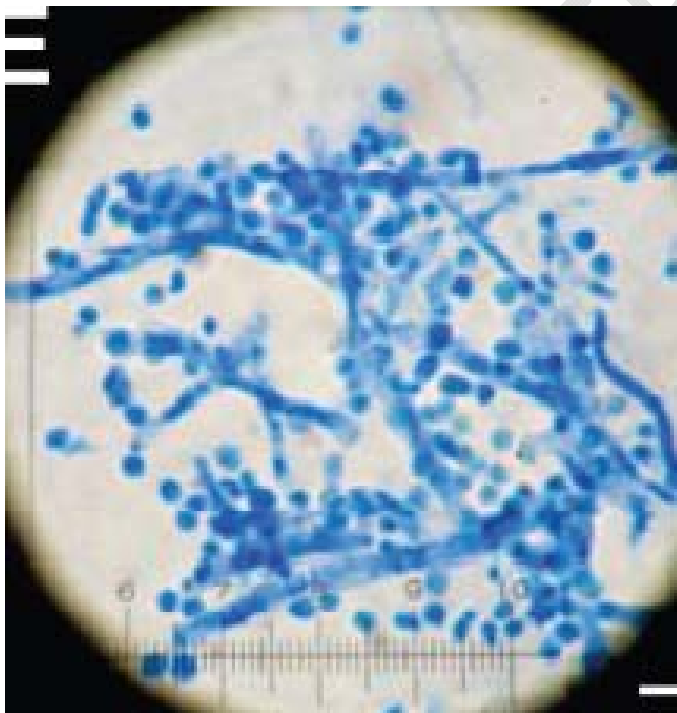
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Fig.9: *Microsporum canis* culture, macroscopic colony



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Fig.10: *Microsporium gypseum* culture, macroscopic colony



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Fig.11: Small conidia (size: 2-3×2-4 μm) and mycelium of *T. mentagrophytes*



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Fig.12: Spiral hyphae of *T.mentoglyphes* var *mentoglyphes* Slide stained with LPCB stain.



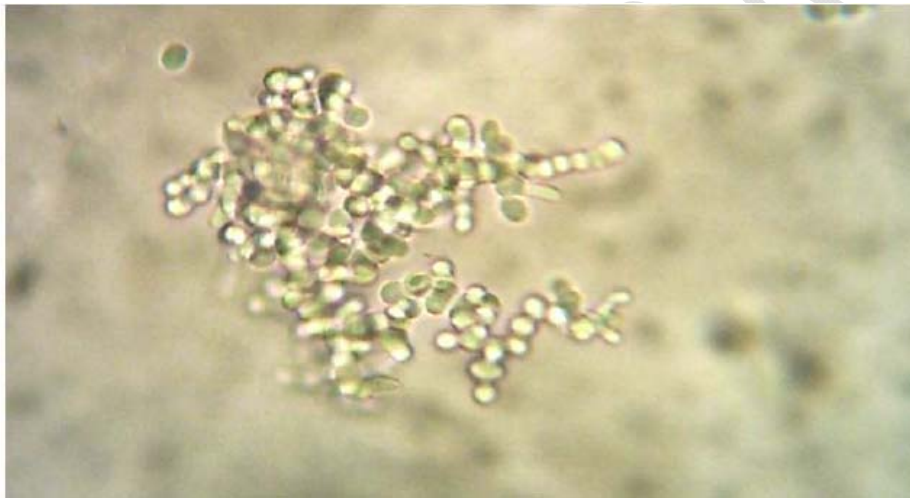
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Fig.13: *Microsporium canis* microscopic observation in lactophenol cotton blue



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Fig.14: *Microsporium gypseum* microscopic observation in lactophenol cotton blue



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Fig.15: Lactophenol cotton blue mount shows chains of chlamyospore of *Trichophyton verrucosum* culture incubated at 37°C.



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172 **Fig.16: Growth of *T.mentagrophytes* on urea agar after 4 days showing hydrolysis of the**
173 **urea.**

174 **Molecular diagnosis:**

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176 Diagnosis with conventional methods is time-consuming because it might take up to 4 weeks or
177 longer to give the final results [35]. Furthermore, morphological identification may be confusing
178 due to polymorphism of dermatophytes [36]. During the last decade, a wide variety of molecular
179 techniques has become available as possible alternatives for routine identification of fungi in
180 clinical microbiology laboratories [37, 38]. *T. mentagrophytes* isolated from nine rabbits and
181 three farm staff were identified by using amplification of CHS-1 gene and ITS+ sequence. The
182 results of sequences of CHS-1 and ITS from different DNA samples revealed that they were
183 identical [21].

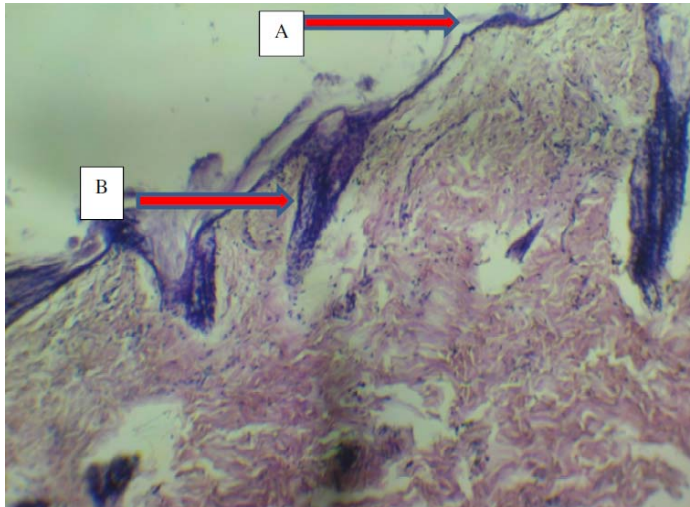
184 **Serodiagnosis:**

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186 Indirect ELISA tests developed to detect specific IgG in rabbits infected with *T. mentagrophytes*,
187 found that (ELISA-rabbits test) is highly sensitive (96.0 %) and highly specific (94.1 %) [39].

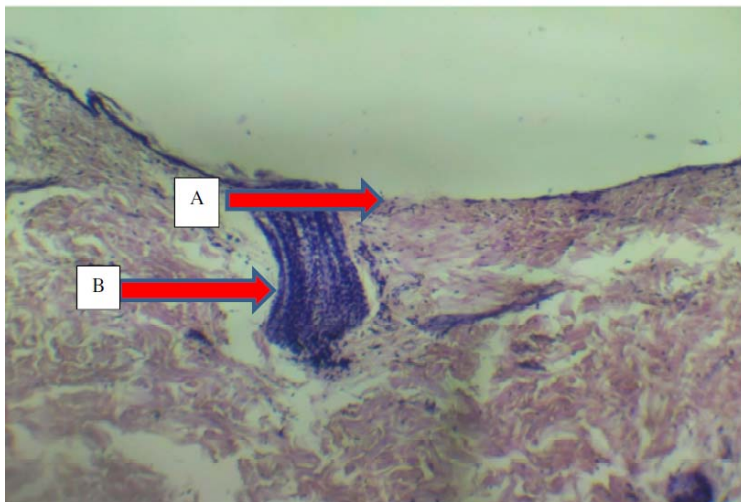
188 **Skin biopsy:**

189 Skin biopsy from rabbit infected with *T. mentagrophytes* showed pathological changes with
190 adherence of fungus to keratinocytes, through the stratum granulosum of the epidermis. In this
191 period of infection there was a hyperkeratosis, thickening of epidermis with hair follicle
192 plugging in addition to keratinized squamous epithelial lining with underlying moderate
193 periappendageal tissue and perivascular chronic inflammatory cells infiltration (lymphocytes)

194 In 8-10 days of induced infection there is keratinized squamous epithelial lining with focal area
195 of surface erosion and underlying moderate periappendageal tissue chronic inflammatory cells
196 infiltration (lymphocytes)



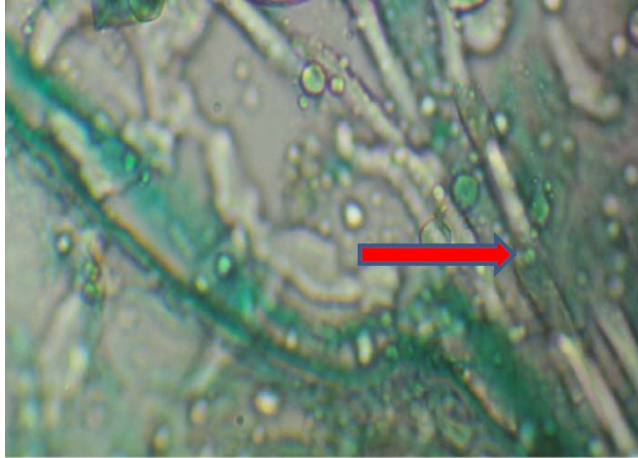
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198 **Fig.17: The bland looking (A) Hyperkeratosis , thickening of epidermis with (B) hair**
199 **follicle plugging in 4-5 days (stained with H&E,10 X)**
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202 **Fig.18: The bland looking with A- surface erosion and B- lymphocytes infiltration in**
203 **8-10 days (stained with H&E,10 X)**
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205 haematoxylin and eosin staining (H&E) may or may not identify dermatophytes and special
206 stains such as periodic acid Schiff (PAS) and Grocott methenamine silver (GMS) are needed The
207 epidermis infiltrated with variable fungal septate hyphae in size in the surface of the squamous
208 epithelium [40].

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212 **Fig.19: Proliferation of septate hyphae of *T. mentagrophytes* in epidermis in 8-10 days**
213 **(stained with PAS,40X).**

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215 **Treatment:**

216 Optimal therapy of dermatophytosis requires a combination of topical antifungal therapy,
217 concurrent systemic antifungal therapy and environmental decontamination. The treatment
218 should be continued until two consecutive negative cultures (at weekly or bi-weekly intervals)
219 are obtained [41]. Topical treatments speed resolution of clinical lesions and may help prevent
220 zoonotic contagion. Systemic therapies that have prolonged residual activity in the skin and hair
221 provide the most effective treatments.

222 **Topical Therapy:**

- 223 1. nystatin ointment for treatment of rabbit experimentally infected with *T. mentagrophytes* for 3
224 weeks [42].
225 2. Clotrimazole is well-documented antifungal agent for treatment of rabbits [43].
226 3. 0.12g of terbinafine 1% cream, for 28 days [44].

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228 **Systemic Therapy:**

- 229 1. Griseofulvin 25–30 mg/kg during 5–6 weeks. Avoid its use in pregnant animals [45,46].
230 2. Itraconazole 5-10 mg/kg daily, for 1 month [47].

231 **Environmental decontamination:**

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233 Enilconazole emulsifiable concentrate will be sprayed onto the walls and ceiling of rabbit house
234 (50 mg per m²) twice weekly for 23 weeks. Treated farm showed reduction of number of
235 clinically infected rabbits [48].

236 **Vaccination:**

237 It is a dried culture of an attenuated strain of *T.mentagrophytes*. It has a high immunogenic
238 activity for dermatophytosis in rabbits. The vaccine is non- reactogenic and is injected
239 intramuscularly. The vaccine has been recommended for practical use in USSR [49].

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241 **Conclusion:**

242 Dermatophytoses are the most common fungal infections in rabbits. Many studies were done
243 considering different aspects of the disease (eg. epidemiology, clinical presentation and
244 diagnosis, treatment, prevention, and control).As many rabbits share the environment with
245 owners as companion animal so they become a source of infection to human this can lead to
246 public health problem.

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