

Original Research Article

Validated Stability – Indicating Methods for Determination of Sofosbuvir by UPLC and HPTLC in Pure Form and Tablet Dosage forms.

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Aims: Two simple and sensitive stability- indicating methods were developed and validated for the quantitative determination of sofosbuvir in presence of its degradation products.

Study design: Ultra high performance liquid chromatography (UPLC), High performance thin layer chromatography (HPTLC) are developed for determination of sofosbuvir in presence of its degradation products, laboratory-prepared mixtures and in tablet dosage forms.

Place and Duration of Study: Analytical Chemistry Department, Faculty of Pharmacy (Girls), Al-Azhar University, between August 2018 and March 2019.

Methodology: Two simple and sensitive stability- indicating methods were developed and validated for the quantitative determination of Sofosbuvir in presence of its degradation products. The first method was an Ultra Performance Liquid Chromatography (UPLC) method, in which efficient separation was carried out on phenomenex kinetex 2.6 μm C18 100 A column using a mobile phase consisting of filtered and degassed mixture of 0.1% ortho-phosphoric acid in water and methanol with the ratio of (40:60% v/v) adjusted to pH 3.5, at a flow rate of 1 mL min⁻¹ and UV detection at 260 nm at ambient temperature. The second method is a high performance- thin layer chromatographic one (HPTLC) in which chromatographic separation was performed on silica gel 60 F₂₅₄ plates, with methanol – chloroform – ammonia (2.5: 6: 1.5 % v/v/v) as a developing system followed by densitometric determination at 261 nm. Sofosbuvir was subjected to stress conditions including alkaline, acidic and oxidative degradation.

Results: Beer's law was obeyed over the range of 1-20 $\mu\text{g mL}^{-1}$ for UPLC and 2-12 μg / spot for HPTC with good accuracy and precision using the two procedures, respectively. Results obtained was statistically analysed and found to be in accordance with those given by the reported method.

Conclusion: The proposed methods were successfully applied for the determination of sofosbuvir in bulk powder, laboratory prepared mixtures and pharmaceutical dosage form with good accuracy and precision. The methods were validated according to ICH guidelines. The results obtained were compared with those of the reported method and were found to be in good agreement.

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13 *Keywords: Sofosbuvir; UPLC; HPTLC; Degradates, Tablet dosage form.*

14 **1. INTRODUCTION**

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16 Sofosbuvir, chemically described as (S)-isopropyl-2-(S)-(2R, 3R, 4R, 5R)-5-(2,4-dioxo-

17 3,4-dihydropyrimidin-1(2H)-yl)-4-fluoro-3-hydroxy-4-methyltetrahydrofuran

18 2-yl)methoxy)phenoxy)phosphoryl amino)propanoate (1), Fig.(1). Sofosbuvir is a direct-

19 acting antiviral agent against HCV for the treatment of adult patients with chronic

20 hepatitis C virus (HCV) infection. It interferes with the HCV lifecycle, restraining

21 viral replication. (2). Literature survey revealed that sofosbuvir has been determined by

22 spectrophotometry and HPLC methods (3-6). It was also assayed in combination with other

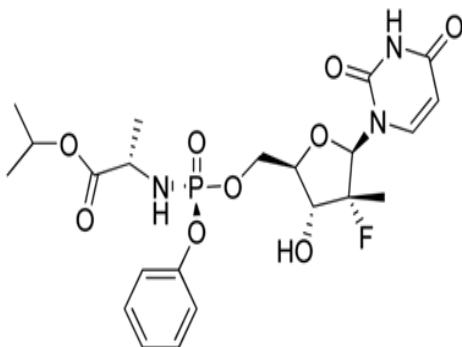
23 drugs by RP- HPLC (7, 8), LC – MS/MS method (9) and bio-analytical UPLC-ESI-MS/MS

24 methods (10-12). Moreover, few stability HPLC indicating methods were reported for its

25 analysis (13-15). The primary goal of this work was to develop a simple, rapid and sensitive

26 methods for the determination of sofosbuvir in presence of its alkaline, acidic and oxidative

27 degradates as well as in its tablet dosage forms.



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Fig. 1: Structure of Sofosbuvir

33 **2. EXPERIMENTAL**

34 **2.1. Instrumentation**

35 - Agelint 1290 Ultra HPLC with binary pump and UV detector (USA).

36 - Camag TLC scanner 3, with WINCATS computer software (Switzerland).

37 - Precoated TLC plates, silica gel 60 GF254 (20 × 20 cm), (Fluka chemie, Switzerland).

38 Using Hamilton 50- μ L micro syringe (Germany).

39 - UV lamp with short wavelength (254 nm), (Desega, Germany).

40 - pH meter combined plus electrode (Adwa model AD1030 pH mv), (UK).

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42 **2.2. Material and Reagents**

43 **2.2.1. Pure and Market samples**

44 Sofosbuvir B.N. OP-G-LD/12/17/099 was kindly supplied by Marcyrl Company for
45 Pharmaceutical Industry, El Obour City, Egypt, with purity of 98.2% as stated by the supplier.
46 MPIVIROPACK® tablets; B. N. 1830486, each tablet labeled to contain 400 mg Sofosbuvir,
47 product of Marcyrl Company for Pharmaceutical Industry, El Obour City, Egypt.

48 **2.2.2. Degraded samples**

49 Ten mg of pure sofosbuvir was accurately weighed and refluxed separately with 25 mL
50 of 5N NaOH for 3 h, or with 25 mL 5N HCL for 7 h. For oxidative degradation 10 mg of pure
51 drug was set aside with 25 mL of 3% H₂O₂ for one week at room temperature. The alkaline
52 and acidic hydrolysed solutions were cooled and neutralized to about pH 7 with 5N HCL or
53 5N NaOH and evaporated to dryness under vacuum. The obtained residues were extracted
54 three times each with 25 mL methanol then filtered separately into 100 mL volumetric flask
55 and diluted to the volume with methanol to obtain a stock solution claimed to contain
56 degradates derived from 0.1mg mL⁻¹ intact drug which was used for UPLC and HPTLC
57 methods. Also oxidative degradate was exposed to dryness at room temperature then the
58 residue was extracted three times each with 25 mL methanol then filtered into 100 mL
59 volumetric flask and diluted to the volume with methanol to obtain a stock solution claimed to
60 contain degradates derived from 0.1mg mL⁻¹ of intact drug which was used for the proposed
61 methods.

62 **2.3. Chemicals and reagents**

63 All reagents used were of analytical grade, solvents were of spectroscopic grade and
64 water used throughout the procedure was freshly distilled.

65 - Methanol; HPLC grade (Sigma-Aldrich, USA).

66 - Ortho phosphoric acid (Fisher, UK)

67 - Sodium hydroxide, hydrochloric acid, chloroform and ammonia (El-Nasr Co., Egypt).

68 **2.4. Standard solutions**

69 Stock solution of the drug (1 mg mL⁻¹) was prepared in methanol. This solution was
70 stable for 1 week in refrigerator or at room temperature. It was diluted with methanol to
71 obtain working standard solution of concentration 0.1mg mL⁻¹.

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73 **2.5. Procedures**

74 **2.5.1. Chromatographic conditions**

75 **i. UPLC method**

76 At ambient temperature, isocratic separation was carried out on Kinetex 2.6μ
77 C₁₈ column 100A (4.6-mm × 100-mm) using a mobile phase composed of mixture of
78 0.1% ortho-phosphoric acid in water and methanol with the ratio of (40:60 % v/v) pH
79 adjusted at 3.5 with ortho phosphoric acid. The mobile phase was pumped at a flow
80 rate 1.0 mL min⁻¹ with an injection volume of 10 μL and the detection at 260 nm.

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82 ii. HPTLC method

83 Analysis was performed on precoated (20 × 20 cm) TLC aluminum silica gel 60 GF254
84 plates. Samples were applied to the plates using Hamilton micro syringe (50 μL). Plates
85 were spotted 2 cm apart from each other and 1.5 cm apart from the bottom edge. The
86 chromatographic tank was pre-saturated with the mobile phase consisting of methanol -
87 chloroform - conc. ammonia (2.5: 6: 1.5 % v/v/v) for 20 minutes, and then developed by
88 ascending chromatography The plates were air dried, and detected under UV- lamp 254 nm.
89 Densitometry was performed at 261 nm in absorption mode. The slit dimension was 6.0 ×
90 0.3 μm and the scanning speed was 20 mm/s with data resolution: 100 nm / step.

91 2.5.2. Linearity

92 i. UPLC method

93 Aliquots of working standard drug solution (0.1mg mL⁻¹) containing (0.01-0.2 mg) were
94 transferred into a series of 10- mL volumetric flasks and diluted to the volume with methanol.
95 10 μL of each solution were injected into the UPLC system in triplicate and
96 chromatographed under the above mentioned conditions. Calibration curve was obtained by
97 plotting the peak area against concentration of the drug.

98 ii. HPTLC method

99 Aliquots of stock standard drug solution (1 mg mL⁻¹) equivalent to (2.0- 12 μg mL⁻¹)
100 were introduced into a series of 10 - mL volumetric flasks diluted to volume with methanol.
101 10μL were spotted on a TLC plate following the above mentioned specific chromatographic
102 conditions and scanned at 261nm. Calibration curve was constructed by plotting area under
103 the peak versus the corresponding drug concentrations in μg / spot.

104 2.5.3. Assay of laboratory prepared mixtures

105 i. UPLC method

106 Different aliquots of intact sofosbuvir working standard solution (0.1mg mL⁻¹)
107 containig (0.01-0.19 mg) were transferred into a series of 10 mL volumetric flasks containing
108 alkaline or acidic or oxidative degradate derived from (0.19 -0.01mg) drug. Ten μL of each
109 solution were injected into the UPLC column and the corresponding chromatograms were
110 recorded at 260 nm; Fig. (2). The intact drug concentrations were calculated from the
111 corresponding regression equation.

112 ii. HPTLC method

113 Different aliquots of stock drug solution (1 mg mL^{-1}) containing (1.0 –11 mg) were
114 introduced into a series of 10 mL volumetric flasks containing (11 –1.0 mg) of the alkaline or
115 acidic or oxidative degraded sofosbuvir and completed to the volume with methanol. Ten μL
116 of each mixture was applied on TLC plate following the procedure under "Linearity"; Fig. (3).
117 Peak areas of the obtained chromatograms were measured and the concentration of the
118 drug was calculated from the corresponding regression parameters.

119 2.5.4. Application to tablet dosage form

120 Ten tablets of Mpiviropack® containing 400 mg of sofosbuvir were powdered and
121 mixed well. An accurately weighed quantity of the powder equivalent to 100 mg was
122 introduced into 100 mL volumetric flask, diluted to volume with methanol and filtered. The
123 obtained solution labeled to contain 1 mg mL^{-1} of the drug was analyzed by HPTLC method.
124 Ten mL of the above solution were diluted to 100 mL with methanol to obtain a solution
125 labeled to contain (0.1 mg mL^{-1}) of the drug analyzed by the proposed UPLC method. The
126 drug concentrations were calculated from the appropriate regression parameters.

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128 3. RESULTS AND DISCUSSION

129 Two chromatographic methods were developed for the selective determination of
130 sofosbuvir in presence of its degradation products by UPLC and HPTLC methods.

131 3.1. Degradation

132 Stressed degradation of sofosbuvir was studied by refluxing the drug using different
133 conditions. Complete degradation was attained upon refluxing the drug with 5N NaOH and
134 5N HCL for 3 h and 7 h respectively at 100°C . Also, oxidative degradation was carried out
135 by keeping of the drug with 3% H_2O_2 for one week at room temperature. The degradation
136 was confirmed with IR using KBr disc and Mass as follow.

137 The pure drug showed appearance of broad band of (OH) group at 3352 cm^{-1} , sharp
138 band of two (NH) group at 3248 cm^{-1} , band of aromatic (CH) group at 3089 cm^{-1} and band of
139 ester (COO) group at 1716 cm^{-1} in its IR spectrum; EI mass showed molecular ion peak at
140 $m/z = 529$ with high intensity (30 %); Fig. (4-a),(5-1). While the alkaline degradate showed
141 appearance of broad band of phosphoric (OH) group at 3421 cm^{-1} in its IR spectrum and
142 disappearance of CH aromatic band at 3009 cm^{-1} , EI mass showed molecular ion peak at
143 $m/z = 453$ with high intensity (19 %), this indicate decreasing in molecular ion peak by 77
144 unit. this means loss of phenyl group. From IR and EI mass analyses, It was concluded that
145 the proposed degradate formed by removal of phenyl group to afford the free phosphoryle
146 group; Fig. (4-b),(5-b).The acidic degradate showed disappearance of the band of C=O
147 group of ester moiety at 1720 cm^{-1} and appearance of broad band of phosphoric (OH) group

148 at 3421 cm^{-1} in its IR spectrum, EI mass showed molecular ion peak at $m/z = 416$ with high
149 intensity (25%), this indicate decreasing in molecular ion peak equal 113 unit It was
150 concluded that the proposed degradate compound formed by removal of isopropyl alaninate
151 moiety to afford the free phosphate group; Fig. (4-c), (5-c). The appearance of ketonic band
152 of (CO) at 1727 cm^{-1} group confirms the oxidation of (OH) group and EI mass showed
153 molecular ion peak at $m/z = 527.15$ with high intensity (32%), this indicate decreasing in
154 molecular ion peak by 2 units; Fig. (4-d),(5-d).
155 Thus a degradation pathway was illustrated in scheme (1).

156 **3.2. Method Development**

157 **3.2.1. UPLC Method**

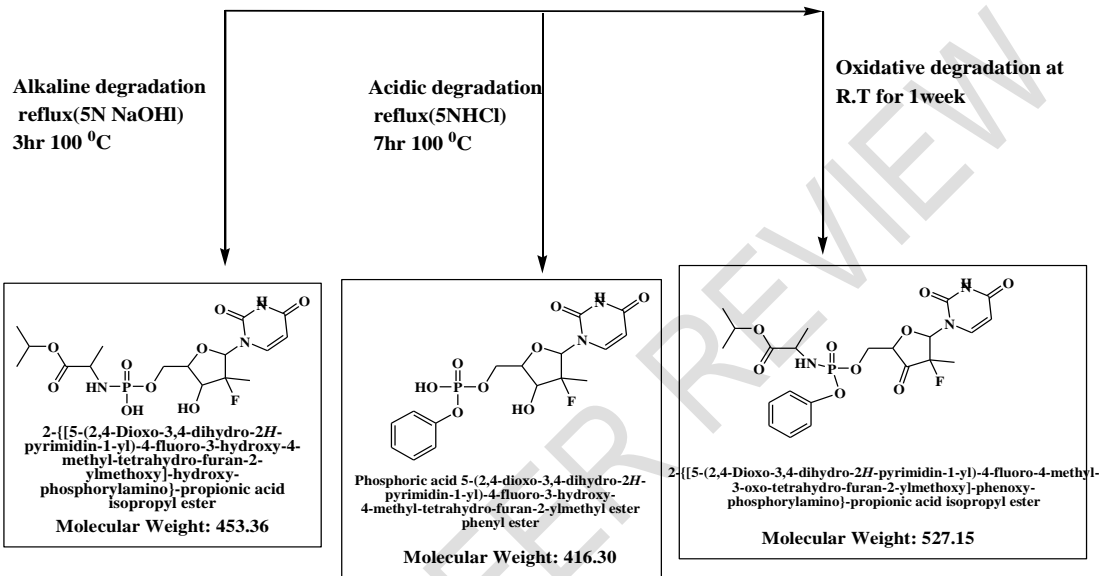
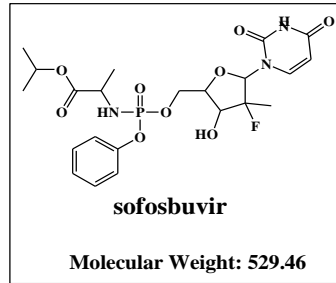
158 Different mobile phases such as ortho-phosphoric acid- acetonitrile -methanol,
159 methanol - water - potassium hydrogen phosphate, acetonitrile - ortho-phosphoric acid,
160 methanol – water and potassium hydrogen phosphate – methanol in different ratios at
161 different pH values were evaluated to obtain optimum resolution. Good resolution with good
162 peak shape and purity were obtained on Kinetex $2.6\mu\text{ C18 100A}$ ($4.6\text{-mm} \times 100\text{-mm}$) column
163 using mobile phase of 0.1% ortho-phosphoric acid in water and methanol (40:60% v/v)
164 adjusted to pH 3.5 with ortho phosphoric . Different flow rates ($0.5\text{ -}2\text{ mL min}^{-1}$) at different
165 wavelengths (250,254, 255, 260, 261, 262 and 265 nm) were tried where optimal flow rate
166 was found to be 1 mL min^{-1} with detection at 260 nm. Intact sofosbuvir was well resolved
167 from the peak of its degradates where R_t was 3.92, 4.27 and 4.15 for alkaline, acidic and
168 oxidative degradates: respectively.

169 **3.2.2. HPTLC Method**

170 Initial studies on the cited drug and its degradation products were carried out to
171 achieve good separation in which different mobile phases in different ratios were tested such
172 as chloroform - methanol- ammonia, chloroform - methanol - acetonitrile, methanol - toluene-
173 acetonitrile, ethyl acetate - methanol - acetonitrile and methanol - water- ammonia and
174 different scanning wavelengths were tried including 250, 245, 255, 260, 262 and 265 nm. It
175 was found that best separation was achieved using a mobile phase of methanol - chloroform
176 - conc. ammonia (2.5: 6: 1.5 % v/v/v). The plates were visualized under UV lamp at 254 nm,
177 where bands appear at R_f 0.73 for intact sofosbuvir and 0.49, 0.38 and 0.64 for its alkaline,
178 acidic and oxidative degradates: respectively. The separated bands of the drug were
179 scanned densitometrically at 261 nm without any interference from its degradates.

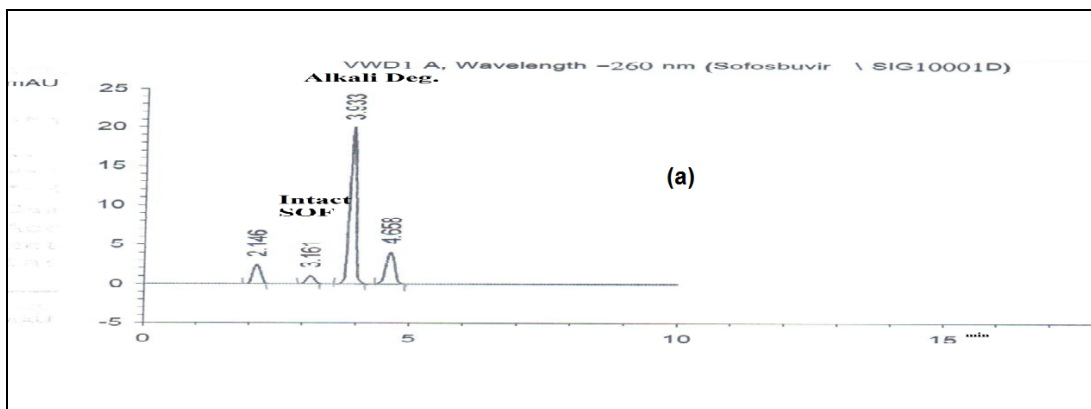
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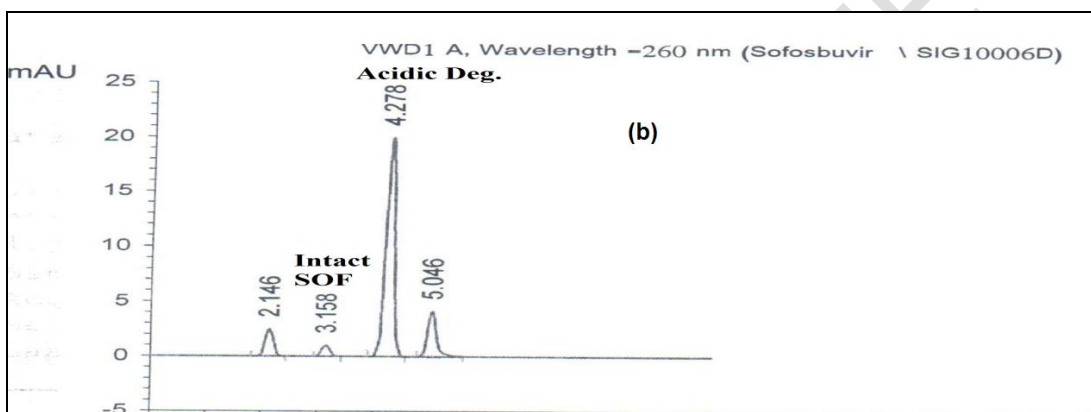
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Chart: 1: Suggested degradation pathway of Sofosbuvir



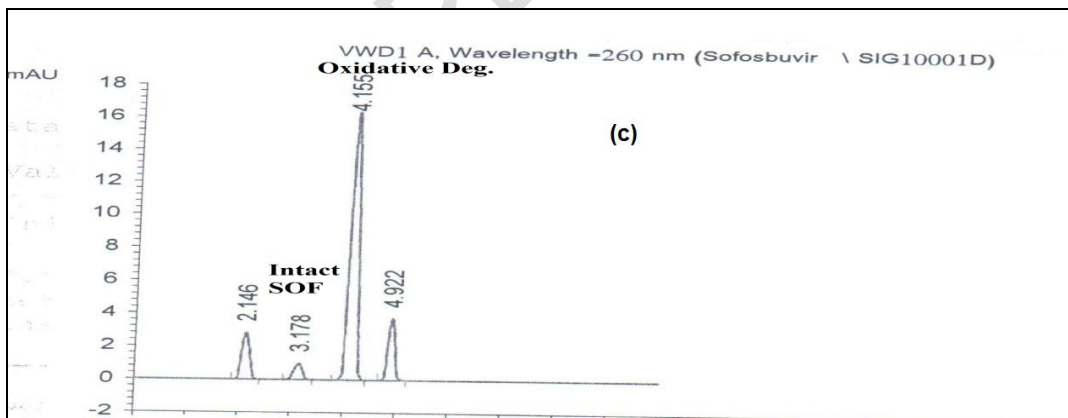
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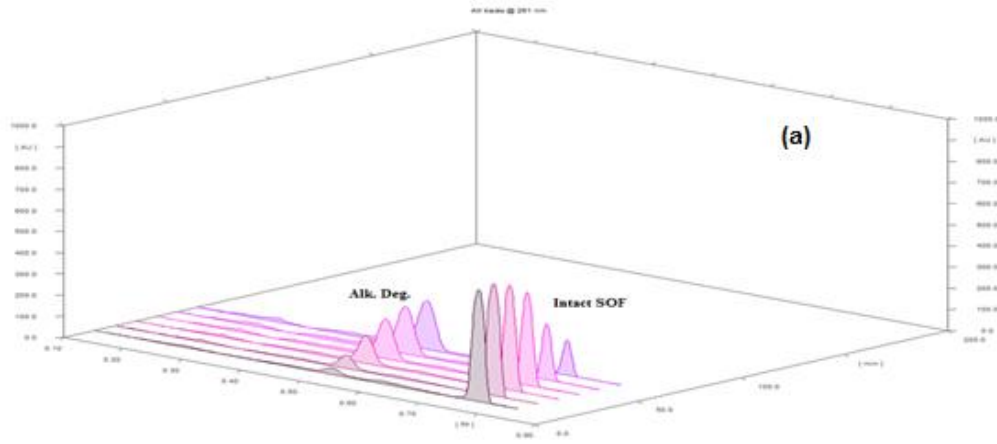
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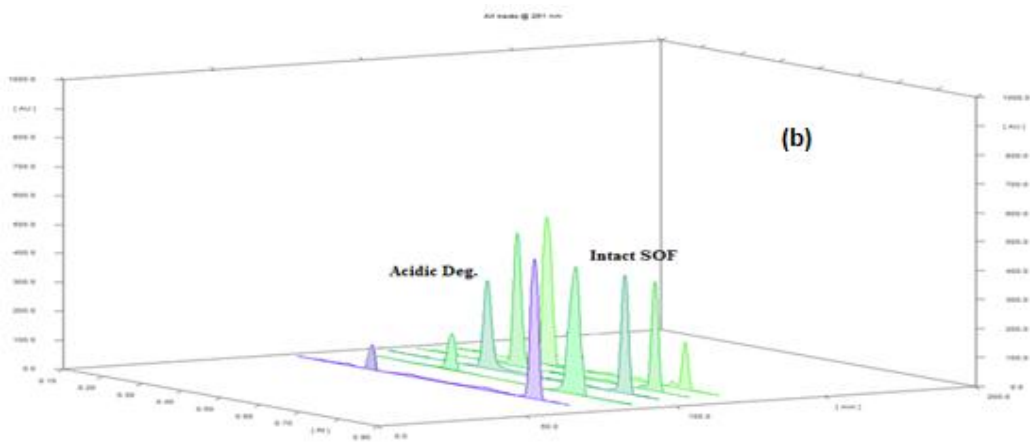
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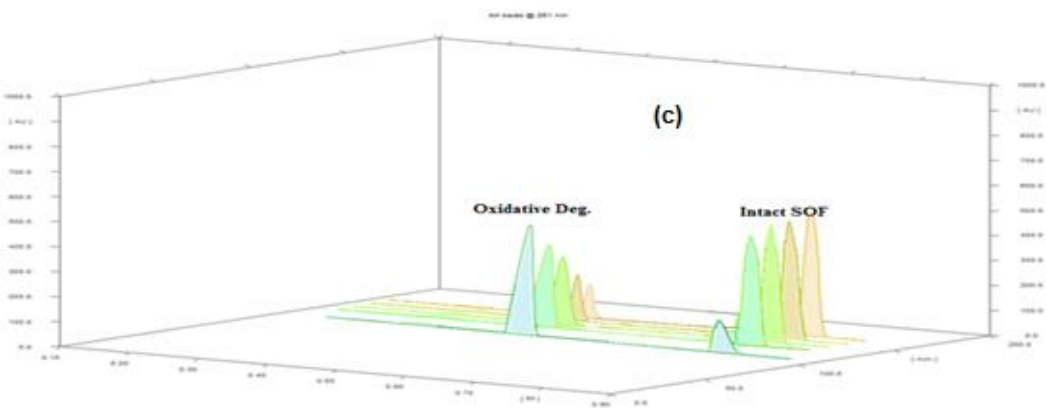
202 Fig. (2): UPLC chromatograms of mixture of ($1 \mu\text{g mL}^{-1}$) of intact Sososbuvir and its
203 alkaline (a), its acidic (b) and its oxidative (c) degradates at 260 nm.



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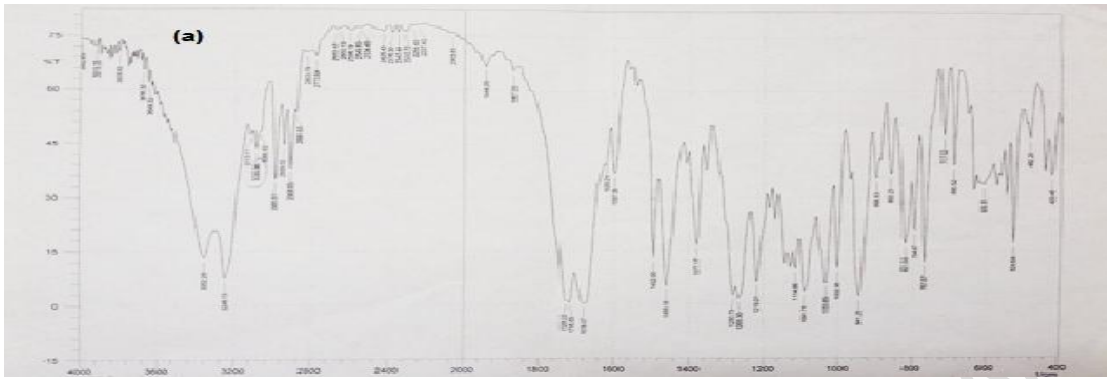
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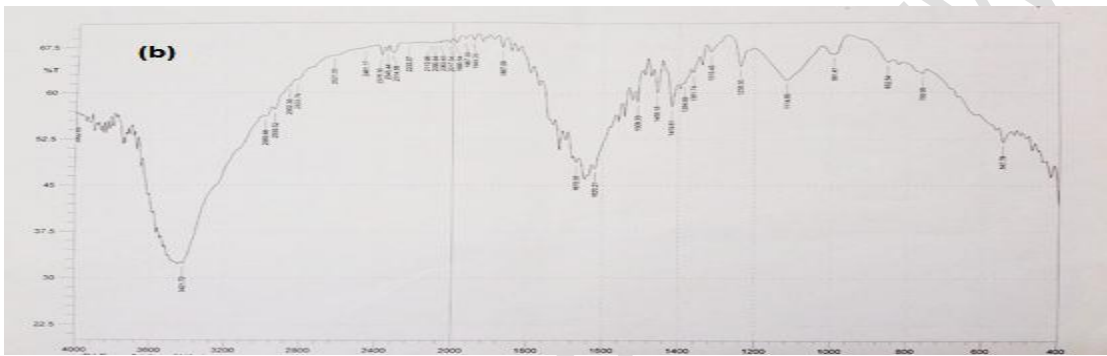
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Fig. (3): Densitometric chromatograms of mixture of intact Sofosbuvir and its alkaline (a), its acidic (b) and its oxidative (c) degradates at 261 nm.

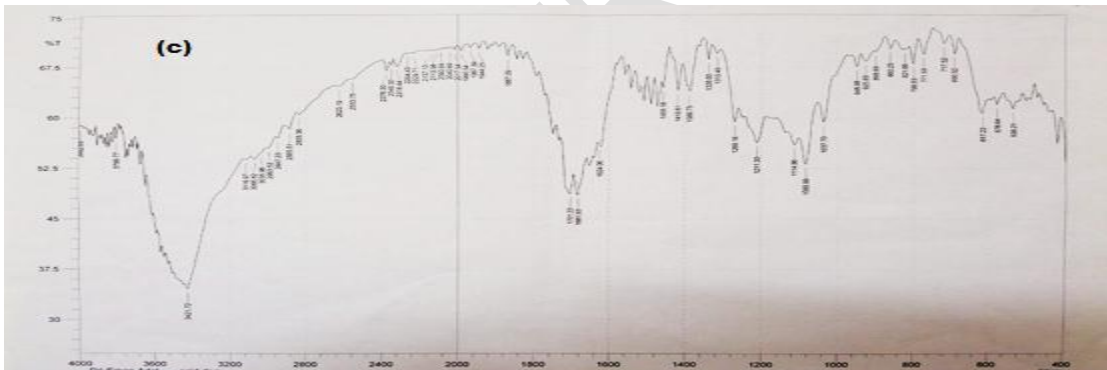
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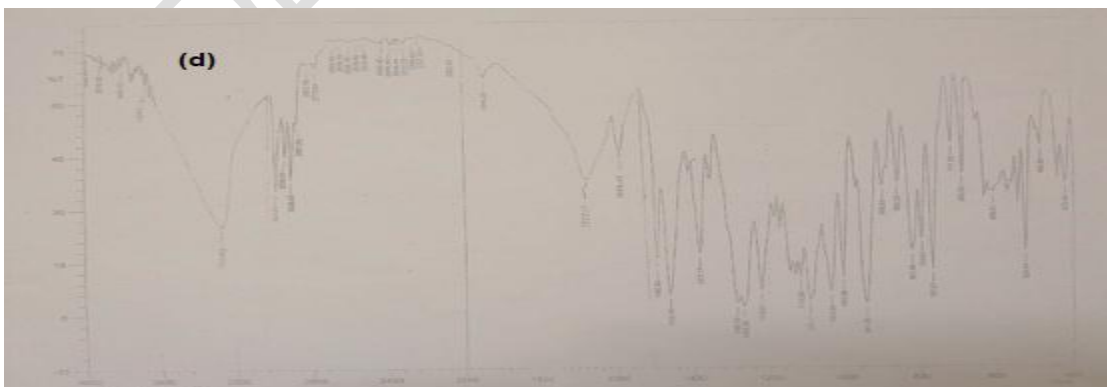
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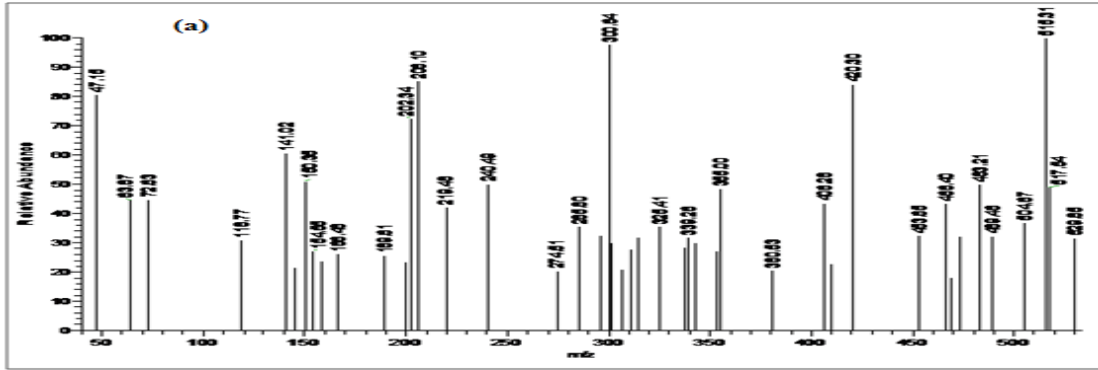


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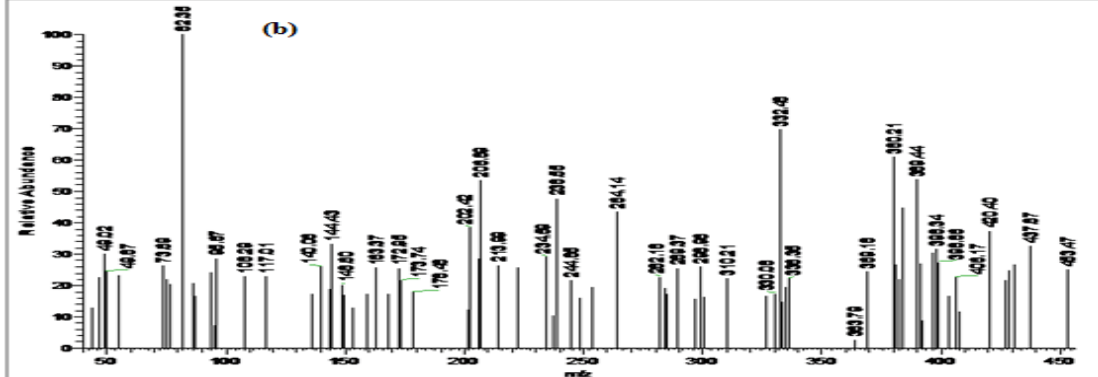


213 **Fig. (4): IR Spectra of intact Sofosbuvir (a), Its alkaline degradate (b), its acidic**
214 **degradate(c),and its oxidative degradate(d) on KBr disc.**

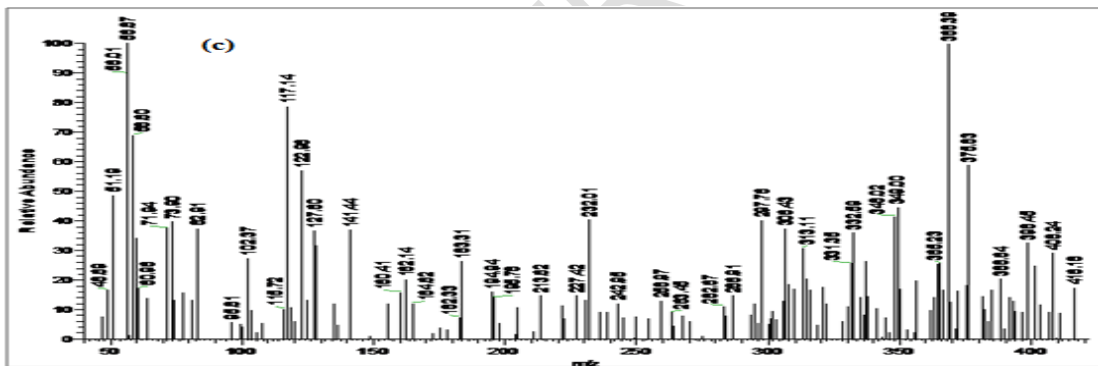
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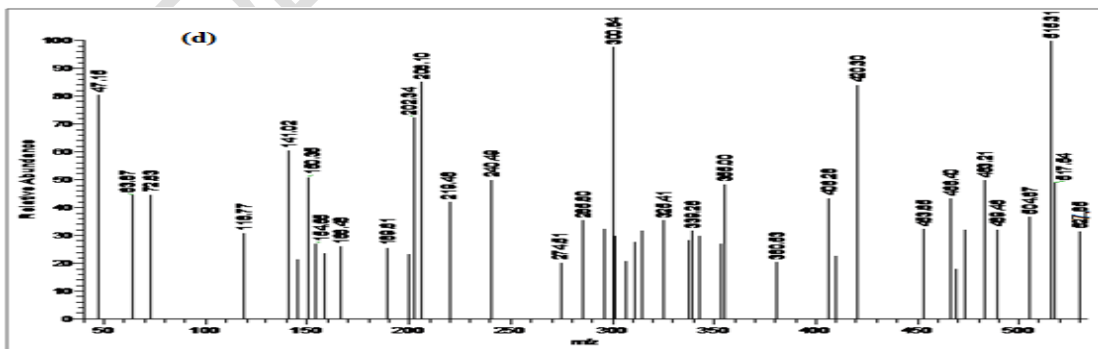
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219 Fig. (5): Mass spectra of intact Sofosbuvir (a), Its alkaline degradate (b), its acidic
220 degradate(c),and its oxidative degradate(d).

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223 3.3. Method validation

224 The methods were validated as per ICH guidelines (16).

- 225 • **System suitability-** System suitability test was performed for UPLC method in
226 accordance with USP. The results shown in Table (1) indicate adequate resolution
227 and reproducibility of the UPLC method and deliberate variations did not affect the
228 system suitability parameters indicate good robustness.
- 229 • **Linearity-** Under the prescribed and experimental conditions a good linearity
230 between peak area and corresponding drug concentration in UPLC and HPTLC
231 methods were obtained over the range of 1-20 $\mu\text{g mL}^{-1}$ and 2-12 $\mu\text{g/spot}$. Regression
232 parameters were computed and presented in Table (2).
- 233 • **Accuracy and precision-** They were assessed by triplicate analysis of three
234 different concentrations covering the linearity range within one day for intraday and
235 three different days for interday analysis. Accuracy (R%) was 99.11% and 100.44%
236 for UPLC and HPTLC methods ; respectively. Intraday precision (RSD %) ranged
237 from (0.68 - 1.66) and (0.60 - 0.63) using the two proposed methods; respectively,
238 was shown in Table (2).
- 239 • **LOD and LOQ-** They were ranged between (0.182, 0.552) and (0.069, 0.210) for
240 both UPLC and HPTLC; respectively, as shown in Table (2).
- 241 • **Selectivity-** It was assured by applying the proposed methods to laboratory
242 prepared mixtures of the intact drug together with its degradation products.
243 Successful selective determination of intact sofosbuvir in presence of up to 95 % of
244 its three degradates in UPLC method as shown in Table (3), while in HPTLC method
245 the selective determination of intact sofosbuvir in presence of up to 91 % of its
246 degradates ; Table (4).
- 247 • **Robustness-**The robustness of the proposed methods was assessed by study the
248 influence of deliberate variation in the mobile phase contents ratio. It was observed
249 that no significant difference in R_f value. The RSD % did not exceed 0.85 and 1.75
250 for both methods; respectively. Also, it checked by studying the effect of different
251 sources of methanol, it was found that using methanol (Sigma – Aldrich, Germany,
252 El-Nasr Co., Egypt) gave RSD% of 0.35 whereas peak area remains acceptable
253 throughout the assay also by checking flow rate in which the produced RSD % was
254 1.32 which proved the robustness of the method.

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258 **3.4. Application to tablet dosage form**

259 The proposed methods were applied for determination of Mpiviropack® Tablets where no
 260 interference from excipients and additives were observed. The results presented in Table (5)
 261 revealed mean recoveries of 98.07% ± 0.89 and 100.73% ± 1.11 for the two proposed
 262 methods; respectively. The recovery of the proposed methods was also validated by
 263 applying the standard addition technique, Table (5). Statistical comparison between results
 264 obtained from the proposed methods and the reported HPLC method (3) for determination of
 265 sofosbuvir in its tablets showed less calculated t and F values than the tabulated ones
 266 revealing no significant difference in accuracy and precision at 95% confidence limit (17);
 267 Table (6). However the proposed methods are much more sensitive and are stability
 268 indicating; determining the intact drug in presence of its three degradates.

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Table 1. Robustness results of system suitability parameters for the determination of sofosbuvir by the proposed UPLC method.

| parameter | Capacity factor (K) | Number of theoretical Plates(N) | Resolution factor (R) | Selectivity factor (α) | Tailing factor (T) |
|---------------------|---------------------|---------------------------------|-----------------------|------------------------|--------------------|
| Mobile phase | | | | | |
| Buffer: MeOH | | | | | |
| mp 39/61 | 2.75 | 6712 | 16.57 | 9.22 | 0.86 |
| mp 40/60 | 2.77 | 6785 | 16.24 | 9.10 | 0.89 |
| mp 41/59 | 2.69 | 6681 | 16.43 | 9.14 | 0.79 |
| pH | | | | | |
| pH 3.6 | 2.80 | 5985 | 16.27 | 8.79 | 0.90 |
| pH 3.5 | 2.73 | 5879 | 16.08 | 8.87 | 0.84 |
| pH 3.4 | 2.70 | 5822 | 16.16 | 8.83 | 0.87 |

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291 **Table 2. Regression analysis and validation parameters for the determination of**
 292 **sofosbuvir by the proposed methods.**

| parameter | UPLC method | HPTLC method |
|-----------------------------------|---------------------------------|------------------------------|
| λ_{max} (nm). | 260 | 261 |
| Linearity range | 1-20 ($\mu\text{g mL}^{-1}$) | 2-12 ($\mu\text{g/spot}$) |
| <u>Regression equation</u> | | |
| slope \pm SD(S_Y) | 8.13 \pm 0.03 | 1364.06 \pm 29.78 |
| Intercept \pm SD(S_X) | -0.21 \pm 0.44 | -79.07 \pm 23.19 |
| SD of residual(S_{YX}) | 0.930 | 248.36 |
| Correlation coefficient (r^2) | 0.9999 | 0.9990 |
| Accuracy (Mean% \pm SD) | 99.11 \pm 0.34 | 100.44 \pm 0.54 |
| Precision (RSD %) | | |
| Intraday precision* | 1.66 | 0.63 |
| Interday precision* | 0.8 | 0.60 |
| LOD | 0.182 ($\mu\text{g mL}^{-1}$) | 0.069 ($\mu\text{g/band}$) |
| LOQ | 0.552 ($\mu\text{g mL}^{-1}$) | 0.210 ($\mu\text{g/band}$) |

293 *Average of 9 determinations.

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**Table 3. Determination of sofosbuvir in laboratory prepared mixtures with its
 degradation products by the proposed UPLC method.**

| Alkaline degradate | | | Acidic degradate | | | Oxidative degradate | | |
|----------------------------------|------------------------------------|---------------------|----------------------------------|-----------------------------------|--------------------|----------------------------------|-----------------------------------|---------------------|
| Intact ($\mu\text{g mL}^{-1}$) | Degraded ($\mu\text{g mL}^{-1}$) | Recovery% of Intact | Intact ($\mu\text{g mL}^{-1}$) | Degradd ($\mu\text{g mL}^{-1}$) | Recover% of Intact | Intact ($\mu\text{g mL}^{-1}$) | Degradd ($\mu\text{g mL}^{-1}$) | Recovery% of Intact |
| 1 | 19 | 100.80 | 1 | 19 | 97.90 | 1 | 19 | 98.30 |
| 5 | 15 | 101.62 | 5 | 15 | 97.40 | 3 | 17 | 99.01 |
| 7 | 13 | 102.85 | 7 | 13 | 99.82 | 9 | 11 | 98.05 |
| 13 | 7 | 101.05 | 13 | 7 | 100.18 | 11 | 9 | 98.50 |
| 15 | 5 | 98.95 | 15 | 5 | 99.06 | 17 | 3 | 101.95 |
| 19 | 1 | 101.15 | 19 | 1 | 97.68 | 19 | 1 | 99.63 |
| Mean% \pm | | 101.07 \pm 1.27 | | | 98.67 \pm 1.18 | | | 99.24 \pm 1.44 |
| SD | | | | | | | | |

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313 Table 4. Determination of sofosbuvir in laboratory prepared mixtures with its
 314 degradation products by the proposed HPTLC method.
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| Alkaline degradate | | | Acidic degradate | | | Oxidative degradate | | |
|--------------------|---------------------|-----------------------|-------------------|---------------------|-----------------------|---------------------|---------------------|-------------------------|
| Intact µg/spot | Degraded µg/spot | Recovery of Intact | Intact µg/spot | Degraded µg/spot | Recovery of Intact | Intact µg/spot | Degraded µg/spot | Recovery % of Intact |
| 10 | 2 | 99.56 | 10 | 2 | 99.92 | 10 | 2 | 97.69 |
| 9 | 3 | 100.43 | 9 | 3 | 100.00 | 9 | 3 | 99.53 |
| 8 | 4 | 101.90 | 7 | 5 | 101.34 | 8 | 4 | 98.51 |
| 7 | 5 | 102.23 | 4 | 8 | 100.41 | 7 | 5 | 99.12 |
| 6 | 6 | 98.69 | 1 | 11 | 101.01 | 1 | 11 | 101.99 |
| 1 | 11 | 101.20 | | | | | | |
| Mean%± SD | | 100.42±1.38 | | | 100.54±0.62 | | | 99.37±1.62 |

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Table 5. Application of standard addition technique for the determination of sofosbuvir in Mpiviropack® tablets by the proposed methods.

| Recovery ±SD% | UPLC Method | | | Recovery ±SD% | HPTLC Method | | |
|------------------|--|---|--------------------------------|------------------|--|---|--------------------------------|
| | Standard addition | | | | Standard addition | | |
| | Claimed taken (µg mL ⁻¹) | Pure added (µg mL ⁻¹) | Recovery % of pure added | | Claimed taken (µg mL ⁻¹) | Pure added (µg mL ⁻¹) | Recovery % of pure added |
| 98.07±0.89 | 5 | 1 | 102.21 | 100.73±1.11 | 4 | 1 | 98.50 |
| | 5 | 5 | 101.99 | | 4 | 3 | 101.02 |
| | 5 | 10 | 99.05 | | 4 | 5 | 100.15 |
| | 5 | 15 | 100.61 | | 4 | 7 | 100.70 |
| Mean%±SD | | | 100.96±1.45 | | | | 100.09±1.12 |

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335 **Table 6. Results obtained by the proposed method compared with reported method⁽³⁾**
 336 **for determination of sofosbuvir in Mpiviropack® tablets.**
 337

| Parameters | Mpiviropack® tablets | | |
|-----------------|--------------------------------|-----------------------------|--------------------------------|
| | UPLC method | HPTLC method | Reported method ⁽³⁾ |
| Linearity range | 1-20 ($\mu\text{g mL}^{-1}$) | 2-12 ($\mu\text{g/spot}$) | 2-60 (μgmL^{-1}) |
| N | 5 | 6 | 5 |
| Mean% | 98.07 | 100.73 | 99.87 |
| SD | 0.89 | 1.11 | 1.60 |
| Variance | 0.79 | 1.23 | 2.58 |
| t- | 2.94(3.18) | 0.66 (3.18) | --- |
| F- | 3.19(9.27) | 2.08(9.27) | --- |

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339 4. CONCLUSION

340 The proposed study describes UPLC and HPTLC methods for estimation of
 341 sofosbuvir in presence of its alkaline, acidic and oxidative degradation products. The
 342 methods were validated and found to be simple, accurate, precise and selective. The two
 343 methods proved their ability to be used for stability indication of the drug. Therefore, they can
 344 be conveniently adopted for estimation, stability studies and routine quality control analysis
 345 of sofosbuvir.

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347 COMPETING INTRESTS

348 Authors have declared that no competing interests exist.

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