Original Research Article

Prevalence of ectoparasitic infection of rodents captured near student's hostels: Zoonotic implications

ABSTRACT

Background and Aims: Rodents constitute more than 42% of the known mammalian species, with 1700 species which belongs to three different families, include Muridae, Microtidae, and Sigmodontidae. Rodents species such as *R. r. diardii* and *R. norvegicus* play an important role as hosts for ectoparasites and reservoirs for various types of viruses, bacteria, rickettsia, protozoa and helminths which are responsible for causing zoonotic diseases to humans and other vertebrate animals. The aim of this work is to identify the species of mites, ticks, and fleas causing diseases to humans and determined the prevalence of infestation in relation to gender, age, and habitat of the rodents.

Place and Duration of the Study: Department of Medical Microbiology and Parasitology, Faculty of Medicine and Health Sciences, University Malaysia, Between September 2018 and March 2019

Methodology: Wild rats were captured using live traps from garbage areas, and places near the cafeteria in the student's residential colleges at University Putra Malaysia. The rats were humanely euthanised and identified. They were classified as adult or juveniles. Their sex was also determined. Ectoparasites were collected by combing the fur the rodents on to a white plan sheet paper. The ectoparasites collected were washed and mounted with Hoyer's media on a glass slide. Parasites were identified using a key morphological feature.

Results: A total of 89 wild rats were trapped and examined for ectoparasites. Eight different species of ectoparasites that comprised of *L. echidnanus*, *L. nuttalli*, *O. bacoti*, *I. granulatus*, *Heamaphysalis* sp., *P. spinoluso*, *H. pacifica*, *X. cheopis*) were identified from the rodents examined. About 55% of the rodents trapped were positive for at least one species of ectoparasite parasites, and about 45.8% of the male rats and 30.8% of female were positive for ectoparasites. Meanwhile, in the adult, 42.9% are positive for at least one species of ectoparasites, whereas 32.2% of the juvenile rodent was also found positive for at least one species of ectoparasites.

Conclusion: The results of this study indicated that rodents trapped from the student's colleges in University Putra Malaysia are infected with various ectoparasites species that they may play an important role in the transmission of certain zoonotic diseases to humans. Therefore, we conclude that there is a potential risk of rodent-borne zoonotic diseases transmission to humans in the study area. Awareness of prevention and control of rodent-borne diseases should be introduced to educate the students on the importance of zoonotic diseases associated with rodents.

Keywords: Ectoparasites, Rodents, Zoonotic, Diseases

1. INTRODUCTION

Several numbers of devastating diseases in tropical and sub-tropical areas of the world are as the result of infection with parasites. A World Health Organization (WHO) report on the leading causes of death worldwide shows that one-third of all deaths are due to parasitic and infectious diseases (32). Rodents play an important role as hosts for ectoparasites and reservoirs for various types of viruses, bacteria, rickettsia, protozoa and helminths which are responsible for causing zoonotic diseases to humans and other vertebrate animals (12). However, these zoonotic diseases from rodents can be transmitted to humans indirectly through ectoparasites such as mites, ticks, and fleas. It can also be transmitted directly through bite wounds, consuming food or water contaminated with rodent feces or urine.

The etiological agents of many infectious diseases utilize invertebrate hosts during their life cycle. Most of these agents are adapted to hematophagous arthropods that share their vertebrate hosts. Therefore, the identification of these arthropod vectors and vertebrate reservoirs is usually a key to sustain an efficient control of vector-borne diseases.

Ectoparasites that include lice, fleas, mites, and ticks are commonly found in wild rats and other rodent species. They are classified into five main groups namely; Mesostigmata (mites), Acarina (ticks), Prostigmata (chiggers), Phthiraptera (lice) and Siphonaptera (fleas) (37). Prevalence studies on ectoparasites infestation in rodent has been reported all over the world (19,17,12, 47, 11, 40, 15, 35, 43, 10, 9). But due to ecological differences in different areas of the country, the parasitic fauna of the rodents in each ecological setting might be different. This notion justifies new studies on parasitic infection of the rodents in other areas of the country.

The present study aimed to determine the prevalence of ectoparasites species in rodents in relation to gender, age, and habitat of the host.

2. MATERIAL AND METHODS

2.1 The study sites

The study was conducted between September 2018 -March 2019 in University Putra Malaysia, which is located (2°59'34.19" N; 101°42'16.79"E) in central Peninsular Malaysia, Kuala Lumpur. The University has seventeen student's residential colleges out of which four colleges were selected randomly for the study. The climate of the study area is tropical climate with, an annual average temperature of about 27-degree Celsius. Typically receives minimum 2,600 mm (100 in) of rain annually; June and July are relatively dry, but even then, rainfall typically exceeds 133 mm (5.2 in) per month.

2.2 Collection of rodents

The rodents were trapped using rectangular metal trap baited with meat, as previously described (4). The traps were set in the late evening closer to garbage storage, students hostel and canteen areas in the University campus. The traps were brought back to the Parasitology laboratory of the Department of Medical Microbiology and Parasitology the following morning for investigation.

2.3 Animal euthanisation

Trapped rodents were euthanised using carbon monoxide. The animals were placed into a sealed chamber, and carbon monoxide was introduced. After the successful euthanisation of the animal, the rodents were removed and place on a clean dissecting board for identification and dissection (48, 24, 4). After the euthanisation, the rats were classified as adult or juveniles based on their weight, length, and the degree of development of their reproductive organs and their gender was also determined (4).

2.4 Collection of ectoparasites

The fur of the animal was combed thoroughly on to a white A4 plan sheet paper using a fine-tooth comb. The Parasites that fell on the white paper from the fur were collected and transferred into a bijou bottle containing 70% alcohol for preservation. Separate containers were used for each animal. A forcep was also used to dislodge the ticks and mites that are difficult to be detached using the comb. The insects collected were preserved before identification (4).

2.5 Mounting and identification of ectoparasites

The preserved ectoparasites were washed using lactophenol and sorted based on their morphology. Preliminary identification of preserved ectoparasites was made under a dissecting microscope. The identification of the ectoparasites was carried out by mounting the parasites on slides with Hoyer's mounting media and observed under a microscope (Nikon eclipse 50í. Japan). The identification was performed by determining the diagnostic characteristics of the ectoparasites. However, the identification of the ectoparasites was based on morphological characteristics using taxonomic keys (28, 44). Identified ectoparasites specimens were classified into four groups, including fleas, mites, ticks, and lice.

2.6 Statistical analysis

All analysis was carried out using graph prism statistical software and excels spreadsheet. Data were presented in percentage. Non-parametric test such as Mann Whitney and Kruskal Wallis test were used to compare the mean differences in parasitic infection between the gender, age, and habitat of the host. P < 0.05 was considered Significance.

3. RESULTS

A total of 89 rodents that comprised of three different species (*R. norvegicus, R. tiomanicus,* and *R. r. diardii*) were examined for ectoparasites. However, eight genera of ectoparasites consisting of seven species that belong to four different groups were identified in the present study. Three species of mites (*L. echidnina, L. nuttalli, O. bacoti*), two species of ticks (*I. granulatus, Heamaphysalis spp.*), two species lice (*P. spinoluso, H. pacifica*), and one species of flea (*X. cheopis*) were identified. Figure 1 shows the prevalence of ectoparasites groups according to host specie. In *R. norvegicus* mites are found to be the most prevalent group with 67.86% prevalent rate, followed by lice and flea which have same prevalence rate of 42.86% respectively, the less prevalent group was tick (41.4%). Ticks are the most encountered ectoparasites in *R. r. diardii* with 45.45% of *R. r. diardii* specie positive for ticks, followed by fleas (34.9%), lice (32.9%) and mites (31.8%). Moreover, in *R. tiomanicus*, fleas (6.71%) are found to the most prevalent ectoparasites followed by ticks (52.94%), lice (32.35%), and mites (23.53%). The overall prevalence of infestation shows that 55% of the rodents captured were positive for at least one species of parasites.

The results in Table 1 show the prevalence and intensity of ectoparasites in relation to the habitat of rodents. All the rodents captured from the four habitats were found infested with similar ectoparasites species. However, rodent trapped from college 11 showed high ectoparasites infestation rate with 45.8% are positive for at least one species of ectoparasites, followed by college 10 with 41%, college chancellor 38%, and college 17 which has the less prevalence rate of 29.6%. A Kruskal-Wallis H test showed that there was a statistically significant difference in the prevalence of ectoparasites infestation between rodents from the four colleges, H value =13.55, P = .0036.

Table 2 shows the distribution of ectoparasites infestation in relation to the gender of the host. The results showed that more male rodents 45.8% were infested with ectoparasites compare to the female that has a prevalence rate of 30.8%. Furthermore, Mann Whitney U test indicated that the differences in terms of parasitic infection between male and female rodent were not statistically significant (P>0.05; *P*=.87). However, in Male rodents, the most prevalent species of ectoparasites species identified were *X cheopis* (64.5%), *H. pacifica* (55.5%), *Haemaphysalis* sp. (50%), *P. spinulosa* (50%). Whereas *I. granulatus* (33.8%), *L. nuttalli* (33.3%), *L. echidnina* (27.7%), *O. bacoti* (27.7%), were found to be less prevalent species. Meanwhile in female *X. cheopis* (37.7%), *P. spinulosa* (35.8%), *O. bacoti* (35.8%), and *H. pacifica* (30.1%) showed the highest prevalent rate, whereas were *L. echidnina* (24.1%), *L. nuttalli* (26.4%), and *I. granulatus* (28.3%) *Haemaphysalis* sp. (28.3%) shows a low infestation rate.

The rodents population was composed of more adult (n=50/56%) than juvenile (n=39/44%). However, both the adult and juvenile wild rats were found positive for ectoparasites parasites. The results show that ectoparasitic infestation was slightly higher among the adult rodents with 42.9% of the adult are infested with at least one species of ectoparasites whereas 32.2% of the male rodent was also found infested with at least one species of ectoparasites, But however the difference was not statistically significant (P = .91, Mann-Whitney U = 30.50, Mean rank for adult = 66.50, Mean rank for juvenile=6950) (Table 3). Among the adult, high infestation rate with *Xenopsylla cheopis* (79.9%), *Polyplax spinulosa* (48.7%), *Haemaphysalis* sp. (46.1%), was observed compared to *L. nuttalli* (38.4%), *L. echidnina* (30.7%), *O. bacoti* (38.4%), *I. granulatus* (33.3%), *H. pacifica* (30.7%). Whereas in juvenile *H. pacifica* 48%, *P. spinulosa* 36%, *Haemaphysalis* sp. 36%, *L. echidnina* (32%), *I. granulatus* (32%), *X. cheopis* (30%), were the most encountered ectoparasites among the juvenile rodents compared to *L. nuttalli* (22%), *O. bacoti* (28%) which are rarely observed.

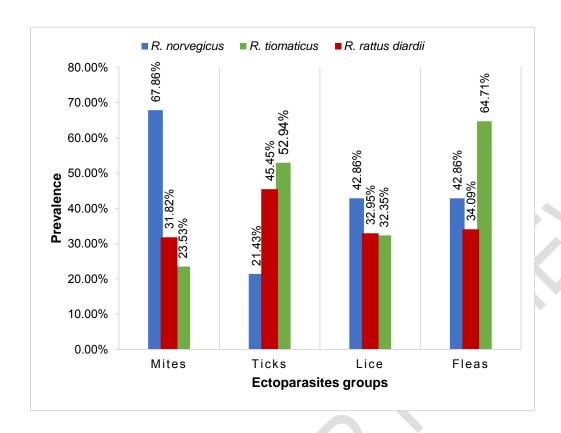


Figure 1: Prevalence of ectoparasites on rodents according species of rodents

Table 1: Prevalence and intensity (Mean \pm SEM) of ectoparasites infection among rodents trapped from student's hostel

Habitat	College 17	College Chancellor (n=19)		College 10 (n=18)		College 11 (n=21)		
No. examined	(n=32)							
	P%	X±SE	P%	X±SE	P%	X±SE	P%	X±SE
L. nuttalli	9 (28.1%)	1.33±0.35	3 (15.7%)	3.00±0.35	7 (33.3%)	1.43±0 .41	7 (38%)	2.57±0.71
L. <mark>echidnina</mark>	9 (28.1%)	1.22±0.74	9 (47.3%)	1.67±0.35	8 (38%)	1.75±0.54	2 (11.1%)	4.00±0.20
O. bacoti	8 (25%)	1.50±0.35	5 (26.3)	1.80±0.35	5 (23.8%)	1.80±0.35	11 (61.1%)	1.00±0.54
I. granulatus	7 (21.8%)	1.29±0.35	6 (31.5%)	1.83±0.54	11(52.3%)	0.91±0.54	11 (61.1%)	1.18±0.74
Haemaphysalis sp.	13(40.6%)	1.23±0.74	7 (36.8%)	1.29±0.35	9 (42.8%)	1.56±0.41	4 (22.2%)	1.50±0.35
P. spinulosa	12(37.5%)	1.17±0.54	7 (36.8%)	1.14±0.20	8 (38%)	1.13±0.35	10(55.5%)	1.20±0.35
H. pacifica	7 (21.8%)	1.29±0.35	10 (52.6%)	1.40±0.74	9 (42.8%)	1.56±0.54	10 (55.5%)	0.80±0.41
X. cheopis	11(34.3%)	1.27±0.74	11 (57.8%)	1.18±0.20	12(57.1%)	1.50±0.35	11 (61.1%)	1.09±0.35
O. P	76 (29.6%)		58 (38%)		69 (41%)		66 (45.8%)	

^{*}Prevalence (P%) = No. of rats infested / Total no. of rats examined multiply by 100, Intensity (mean abundance = Mean no. of ectoparasites collected / Total no. of host infested.

Table 2: Distribution of the ectoparasites infection according to the gender and species of the host

Ectoparasite species	Male (n=36)				Female (n=53)			
	RR	RN	RT	Total	RR	RN	RT	Total
L. nuttalli	5 (13.8%)	4 (11.1%)	3 (8.3%)	12 (33.3%)	6 (11.3%)	5 (9.43%)	2 (3.77%)	14 (22%)
L. echidnina	8 (22.3%)	3 (8.33%)	4 (11.1%)	15 (41.6%)	4 (7.54%)	7 (13.2%)	1 (1.88%)	13 (32%)
O. bacoti	4 (11.1%)	4 (11.1%)	2 (5.5%)	10 (27.7%)	10 (18.8%)	7 (13.2%)	2 (3.77%)	19 (28%)
Ticks								
I. granulatus	6 (16.6%)	4 (11.1%)	4 (11.1%)	14 (38.8%)	7(13.2%)	5 (9.43%)	3 (5.66%)	15 (32%)
Haemaphysalis sp.	10 (27.7%)	6 (16.6%)	2 (5.5%)	18 (50%)	8 (15.0%)	5 (9.43%)	2 (3.77%)	15 (36%)
Lice								
P. spinulosa	9 (25.0%)	5 (13.8%)	4 (11.1%)	18 (50%)	7 (13.2%)	7 (13.2%)	5 (9.43%)	19 (36%)
H. pacifica	8 (22.3%)	10 (27.7%)	2 (5.5%)	20 (55.5%)	7 (13.2%)	5 (9.43%)	4 (7.54%)	16 (48%)
Flea								
X. cheopis	12 (33.3%)	9 (25.0%)	4 (11.1%)	25 (69.4%)	9 (16.9%)	7 (13.2%)	4 (7.54%)	20 (30%)
O. P				45.89	%			30.8%

^{*}RR = R.r. diardii, RN = R. norvegicus, RT = Rattus tiomanicus, RT = Rattus tiomanicus,

Table 3: Distribution of the ectoparasites infection according age of the host

Ectoparasite species	Adult	Juvenile (n=39)		
	(n=50)			
Mites				
L. nuttalli	15(38.4%)	11(22%)		
L. echidnina	12(30.7%)	16(32%)		
O. bacoti	15(38.4%)	14(28%)		
Ticks				
I. granulatus	13(33.3%)	16(32%)		
Haemaphysalis sp.	18(46.1%)	15(36%)		
Lice				
P. spinulosa	19(48.7%)	18(36%)		
H. pacifica	12(30.7%)	24(48%)		
Flea				
X. cheopis	30(79.9%)	15(30%)		
Overall prevalence	42.9%	32.2%		

4. DISCUSSION

The ectoparasites of rodents play an important role as vectors of pathogenic microorganisms that transmit different diseases to humans (20,29). However, several studies on ectoparasites of rodents were reported in Malaysia (25,33,39), and other neighboring countries, including Indonesia, Viet Nam, Bangladesh, and Singapore. (11,) Fleas are known to transmit bubonic plague, essentially a zoonotic disease caused by a bacteria *Yersinia pestis*, from rodents to humans. However, in South and Southeast Asia, plague remains endemic in several regions (e.g., India and Viet Nam), with regular outbreaks among humans (11). The flea species *X. cheopis* and *C. canis* serve as intermediate hosts for species of tapeworms that occasionally infect humans (38).

Furthermore, it has been reported that fleas serve as vectors of several diseases that include Salmonellosis, Tularemia, Leishmaniasis, Trypanosomiasis and relapsing fever infections. In the current study, we recorded the presence of *X. cheopis* among wild rat captured at the prevalence rate of 84.4%. However, the high prevalence of *X. cheopis* in the present study indicates a potential risk of transmission of diseases (Rickettsia typhi and plague,) associated with this parasite to humans. Other previous studies have reported a high prevalence of *X. cheopis* in rodents (25, 33,9,34). According to WHO in 2015, the *X. cheopis* on wild rats represents a potentially dangerous situation with regard to increased plague risk for human beings in the event of an outbreak of plague (WHO Plague Manual). Although, there have not been outbreaks in recent years in Peninsular Malaysia. But, It's endemic in other Southeast Asian countries, including Indonesia (13,45). Thailand (8), Vietnam (36) and Myanmar (3). The first case of plague in Malaysia occurred

in Penang in 1896, and the most recent case was in Perak in 1928 (30). The mite species *L. nuttali, L. echidinus* and *O. bacoti* are also ectoparasites that are often found in rodents. However, in the present study, all three species of mites were found in the trapped rodents. The *L. nuttali, L. echidnina* and *O. bacoti* are medically importance ectoparasites species, and they are known to transmit diseases to humans. The mite species *O. bacoti* does not only parasitizes the wild and domestic rats but also bites the human as the accidental host, consequently leading to transmission of filariasis to humans since that the *O. bacoti* is the intermediate host of the filarial worm (40).

Furthermore, *O. bacoti* also caused mite dermatitis in humans. The first case report of *O. bacoti* causing dermatitis in humans was reported from Australia, followed by other cases reported in the USA and Germany (2). It has been estimated that approximately 80% of the wild rodents in Germany are infested by this parasite (2). In Malaysia, the first authentic case of dermatitis caused by *O. bacoti* was reported in 1974 (31). Therefore, the presence of *L. nuttali, L. echidnina*, *O. bacoti* in the rodent's population in the student's residential colleges in UPM may have a potential risk to students living in the respective colleges.

Two species of ticks recovered in the present study include; *Haemaphysalis* sp., and *I. granulatus*. The *Haemaphysalis* sp. is medical importance tick species which transmits different groups of pathogens including protozoa (Babesia), bacteria (Tularemia), Richettsia sp. and arboviruses. Their bites can also cause stress and blood loss to the animal and human hosts (1). *I. granulatus* is also one of the medical importance ectoparasites because it is the main vector of Langat Virus (42). Apart from Langat virus, *Ixodes* sp. is also known to transmit other pathogens such as babesiosis, human granulocytic anaplasmosis, Lyme disease (28). In Malaysia, *Ixodes* sp. is involved in the spread of tick typhus and Q fever to humans in the climax forest of Peninsular Malaysia (26). The high prevalence of tick species (77.7%) infesting rodents seen in the present study may be due to the suitable environment for tick survival, because ticks are likely to be found in habitats such as shrubs, forest, and plantation and some of the students hostels in UPM where the trapping of rodents conducted was very close to forest plantations. Previous similar studies have also reported these tick species infesting wild rats and other forest rodents (5,12,18,27).

The *P. spinulosa* and *H. pacifica* were two common lice species encountered in the present study. They were found infecting rodent population at the prevalence rate of 37%, and 36 % respectively. Lice are medically importance ectoparasites in both human and rodents; they are known to harbor and transmit plague bacilli and transmit tularemia bartonellosis to humans. There bites in human causes a condition called pediculosis. Furthermore, *P. spinulosa* can also transmit *T. lewisi*, *Hemobartonella muris*, and Rickettsia typhi and the clinical manifestation associated with *Polyplax* sp. comprised of anaemia and general unthriftiness, leading to debilitation (5). In Malaysia, infestation with *Polypolax* sp and *H. pacifica* has been previously reported in urban rats (37,33). *H. pescinata*, *P. spinulosa* and *H. pacifica* have also been previously reported in rodents from Kuala Selangor Nature Park (5). Similar findings on rodents ectoparasites infestation with louse species *P. spinulosa* and *H. pacifica* has been reported worldwide (10, 47, 41, 15).

Age-related differences in term of ectoparasitic infestations among the wild rats observed in the present study could be attributed to the fact that older or adults rats have a longer exposure time to potential infection compared to juvenile (7). On the other hand, the low prevalence of infestation observed in the juvenile rats is probably biased due to the low number of juvenile rats captured in the present study. Other finding with similar observations for host age differences in infestation were also reported (37).

More male (45.8%) rats were infested with ectoparasites despites there low number compare to the female which has a prevalence rate of 30.8%. However, this finding may be as a results of wider home range of male rats that tend to overlap (in search for food, reproductive partner, courtship) which increases their exposure to ectoparasitic infections, whereas the reproductive female shows a stronger site-specific organization which could explain the low rate of transmission (21,6). Other previous studies have reported a similar finding (21,22). The similarities in species recovered from four different locations may be as a result of similarity in the geographical structure of the samplings sites since the trapping was carried out within similar geographical areas that have similar vegetation cover. It has been previously reported similarity in geographical structure can bring the similarity in the fauna of the ectoparasites in the different regions (22).

4. CONCLUSION

The finding of this study showed that wild rats captured from four colleges (College 17, College Chancellor, College 10 and 11) in UPM are infected with different ectoparasites of zoonotic importance. This suggests a potential risk of arboborne disease transmission to humans. Therefore, there is a need Awareness of prevention and control of rodent-borne diseases to educate the students on the importance of zoonotic diseases associated with rodents. Moreover, further studies should focus on the distribution of diseases which are transmitted by ectoparasites to humans not only in UPM campus but also in places, where most of the population live in close contact to rodents, livestock, and dogs.

ETHICAL APPROVAL

All authors hereby declare that Principles of laboratory animal care (NIH publication No. 85-23, revised 1985) were followed, as well as with guidelines of the animal care and use committee (ACUC), University Putra Malaysia. All experiments have been examined and approved by the Animal ethics committee of the University of Putra Malaysia (Ref. No: UPM/IACUC/AUP-R039/2018).

REFERENCES

- 1. Louis AM. Global importance of ticks and associated infectious disease agents. Clinical Microbiology Newsletter. 2009;31(5):33-36
- 2. Beck W, Folster-holst R. Tropical rat mites (*Ornithonyssus bacoti*) serious ectoparasites. Journal of German Society of Dermatology. 2009;(7):667-670.
- 3. Brooks JE. Plague in small mammals and humans in Rangoon, Burma. Southeast Asian Journal of Tropical Medicine and Public Health. 1977;8(3):335-344.
- 4. Herbreteau V, Jittapalapong S, Rerkamnuaychoke W, Chaval Y, Cosson JF. and Morand S, editors. Protocols for field and laboratory rodent studies. 2th ed. Kasetsart University press 2011.
- 5. Chuluun B, Mariana A, Ho T, Mohd K. A preliminary survey of ectoparasites of small mammals in Kuala Selangor Nature Park. Tropical Biomedicine. 2005;22(2):243-247.
- 6. David E, John T, Emlem J R. Studies on home range of brown rats. Journal of Mammalogy. 1948;29(4):207-225.
- 7. Easterbrook JD, Kaplan JB, Glass GE, Watson J, Klein SL. A survey of rodent-borne pathogens carried by wild-caught Norway rats: A potential threat to laboratory rodent colonies. Laboratory Animals. 2007;42(1):92-98.
- 8. Elbel, RE, Thaineua MA. Flea and Rodent Control Program for Plague Prevention in Thailand. The American Journal of Tropical Medicine and Hygiene. 1957;6(2):280-293.
- 9. Fagir DM, El-Rayah EA. Parasites of the Nile rat in rural and urban regions of Sudan. Integrative Zoology. 2009;4(2):179-187.
- 10. Frye M J, Firth C, Bhat M, Firth M, Che X, Lee D, Williams SH. Preliminary survey of ectoparasites and associated pathogens from Norway rats in New York City. Journal of Medical Entomology. 2015;52(2):253-259.
- 11. Fuehrer HP, Igel P, Treiber M, Baumann TA, Riedl J, Swoboda P, Noedl H. Ectoparasites of livestock, dogs, and wild rodents in the Chittagong hill tracts in Southeastern Bangladesh. Parasitology Research. 2012;111(4):1867-1870.
- 12. Gholamhossein M, Hamidi K, Nourani L. Occurrence of ectoparasitic arthropods (Siphonaptera, Acarina, and Anoplura) on rodents of Khorasan Razavi Province, northeast of Iran. Asian Pacific Journal of Tropical Disease.2015;18(8):7-12.
- 13. Hadi TR, Sarbini SD. Survey of small mammal ectoparasites in West Sumatra, Indonesia. Southeast Asian Journal of Tropical Medicine and Public Health. 1981;12(2):7-27.
- 14. Han BA, Schmidt JP, Bowden SE, Drake JM. Rodent reservoirs of future zoonotic diseases. Proceedings of the National Academy of Sciences.2015;112 (22); 7039-7044.
- 15. Harrison A, Robb GN, Alagaili AN, Hastriter MW, Apanaskevich DA, Ueckermann EA, et al. Ectoparasite fauna of rodents collected from two wildlife research centres in Saudi Arabia with discussion on the implications for disease transmission. Acta Tropica. 2015;147(3):1-5.
- 16. Jongejan F, Uilenberg G. The global importance of ticks. Parasitology.2015;129(11):3-14.
- 17. Sumangali RP, Rajapakse RS. Ecto and Endo Parasites of rodents from two selected sites in Kandy district. Ceylon Journal of Science. 2012;41 (1):71-77.
- 18. Kim CM, Yi YH, Yu DH, Lee MJ, Cho MR, Desai AR, Chae JS. Tick-borne rickettsial pathogens in ticks and small mammals in Korea. Applied and Environmental Microbiology. 2006;72(9):5766-5776.
- 19. Kohls GM. Malaysian Parasites-XVIII. Ticks (Ixodoidea) of Borneo and Malaya. Studies from the Institute for Medical Research, Federated Malay States. 1957:28
- 20. Kourosh A, Mitra S, Iraj M, Amir A, Hamid H, Mohammad A, et al. Intestinal Helminths in different species of rodents in North Khorasan Province, Northeast of Iran. Iranian Journal of Parasitology. 2017;12 (2):267-273.
- 21. Kowalski K, Bogdziewicz M. Sex differences in flea infections among rodent hosts: is there a male bias? Parasitol Res. 2015; 114(9):337-341
- 22. Krasnov BR, Stanko M, Matthee S, Laudisoit A, Leirs H, Khokhlova IS, Serge V. Male hosts drive infracommunity structure of ectoparasites. Oecologia, 2011;166:1099-1110.
- 23. Cooperation health collaborative project. Biology and Control of Vector-Borne Infections in Europe. Large-Scale Integrating Project, 2010:1-195.
- 24. Leary S, Underwood W, Anthony R, Cartner S. Guidelines for the euthanasia of animals: 2nd ed. American Veterinary Medical Association. 2013

- 25. Madinah A, Fatimah A, Mariana A, Abdullah MT. Ectoparasites of small mammals in four localities of wildlife reserves in Peninsular Malaysia. Southeast Asian journal of tropical medicine and public health. 2011;42(4):803-813.
- 26. Marchette. Rickettsioses (tick typhus, q-fever, urban typhus) in Malaya. Journal of Medical Entomology.1966; 2(4): 339-371.
- 27. Mariana Z, Zuraidawati TM, Mohd KI, Saleh, MS. A Survey of Ectoparasites in Gunung Stong forest reserve, Kelantan, Malaysia. Southeast Asian Journal of Tropical Medicine and Public Health.2005; 36(5):1125-1131.
- 28. Mathison BA, Pritt S. Laboratory Identification of arthropod ectoparasites. Clinical Microbiology Reviews. 2014;27(1):48-67.
- 29. Meerburg BG, Singleton GR, Kijlstra A. Rodent-borne diseases and their risks for public health. Critical Reviews in Microbiology. 2009;35(3):221-270
- 30. Milne JC. A brief review of fifty years of medical history in Selangor, Federation of Malaya. Medical Journal of Malaya. 1948;2(3):161-173.
- 31. Nadchatram M, Ramalingam S. Dermatitis caused by *Ornithonyssus bacoti*. Southeast Asian Journal of Tropical Medicine and Public Health.1974;5(1):15.
- 32. World Health Organization report (WHO). 50 Facts: Global health situation and trends.1955-2025. World health report. 2018. Accessed 3 September 2018. Available: https://www.who.int/whr/1998/media centre/50facts/en/
- 33. Nursheena S, Mohd-Zain SN, Arnez S, Khalil S, Braima KA, Abdulaziz NM., et al. Ectoparasites of murids in Peninsular Malaysia and their associated diseases. Parasites and vectors. 2015;8(2);1-10.
- 34. Ogunniyi T, Balogun H, Shasanya B. Ectoparasites and Endoparasites of Peridomestic House-Rats in Ile-Ife, Nigeria and Implication on Human Health. Iranian Journal of Parasitology.2014; 9(1):134-140.
- 35. Okorafor KA, Odaibo AB, Eleng I, Okete JA. Occurrence and prevalence of ecto and gastrointestinal parasites in Wild cane Rats (*Tryonomys Swinderianus*) from Oyo State, South-Western Nigeria. European Journal of Zoological Research.2012;1(3):70-76.
- 36. Olson WP. Rat-flea indices, rainfall, and plague outbreaks in Vietnam, with emphasis on the Pleiku area. The American Journal of Tropical Medicine and Hygiene. 1969;18(4):621-628.
- 37. Paramasvaran S, Sani RA, Hassan L, Krishnasamy M, Jeffery J, Oothuman P, et al. Ectoparasite fauna of rodents and shrews from four habitats in Kuala Lumpur and the states of Selangor and Negeri Sembilan, Malaysia and its public health significance. Tropical Biomedicine. 2009; 26(3): 303-311.
- 38. United State Department of Health, Education, and Welfare public health service Communicable Disease Center. Fleas of public health importance and their control. Public Health Service Publication. 1962;772(5):1-10
- 39. Premaalatha B, Nurulaini R, Zawida Z, Norakmar I, Zaini CM, Ramlan M. A Survey of bacterial and parasitic infections of rats caught in the veterinary research institute, Ipoh. Malaysian Journal of Veterinary Research. 2010;6(7):45-50.
- 40. Rahdar M, Vazirianzadeh B. Identification of collected ectoparasites of rodents in the west of Khuzestan Province (Ahvaz and Hovizeh), Southwest of Iran Asian Pacific Journal of Tropical Disease. 2015;1808(4):1-6.
- 41. Shahi M, Pakari A. A study on rodent ectoparasites in Bandar Abbas: The main economic southern seaport of Iran. Iranian Journal of Environmental Health Science & Engineering. 2007;4(1):173-176.
- 42. Smith CE. A virus resembling Russian spring-summer encephalitis virus from an ixodid tick in Malaya. Nature. 1956;(4):581-582.
- 43. Stanko M, Miklisova D, De Bellocq JG, Morand S, Mammal density and patterns of ectoparasite species richness and abundance. Oecologia. 2002;131(2):289-295.
- 44. Anne M. Zajac Gary A. Conboy. Veterinary Clinical Parasitology. 8th ed. UK: Wiley Blackwell:2012
- 45. Turner RW, Martoprawiro S. Dynamics of the plague transmission cycle in central Java (ecology of potential flea vectors). Health Studies in Indonesia.1974;11(2).
- 46. World Health Organization (W.H.O). Report on Global Surveillance of Epidemic-prone Infectious Diseases. Accessed 2 March 2018.

 Available http://www.who.int/csr/resources/publications/plague/CSR ISR 2000 1/en/index5.html
- 47. Yang P, Oshiro S, Warashina, W. Ectoparasitic Arthropods occurring on *Rattus norvegicus* and *Rattus rattus* collected from two properties on the Island of Oahu, Hawaii (Acarina, Siphonaptera, and Anoplura). Proceeding of Hawaiian Entomological Society. 2009; 41:53-56
- 48. Lim Boo Liat, The house rodents and house shrew in Malaysia and Southeast Asia. Utar Agriculture Science Journal 2015;1(4) 2015:43-50