Bacteriological Quality of Kunu Drink Sold In Bayelsa State Nigeria and the Pathogenic Potential of Some Isolates.

ABSTRACT

Introduction: Kunu or kununzaki is a beverage drink made from grains such as millet, sorghum and maize or other combinations. It is a non-alcoholic beverage marketed in public places such as offices, markets, schools, motor parks and used in festivities such as weddings, birthday celebration, naming ceremonies etc. The high bacterial content of kunu calls for investigation.

Aim: The aim of this research is to isolate, identify bacterial contaminants in kunu and determine enterotoxin producing abilities of some isolates.

Methodology: A total of 150 bottles of kunu were purchased, 50 each from Yenagoa, Sagbama and Ogbia respectively. Each bottle of kunu was properly mixed by gentle inversion several times and 1mL of the kunu was pipetted and added to 9mL sterile peptone water. Subsequent serial dilution was made to 10^-5. Then 0.1mL was placed on agar media in duplicate. The plates were incubated at 37°C for 18-24 hours and examined for growth.

Results: The bacteria isolated from Kunu were S. aureus 150 (27.8%), E. coli 150 (27.8%), Bacillus sp. 150 (27.8%) and Staphylococci sp. 90 (16.7%) respectively. Out of the S. aureus isolated, 25 (16.6%) produced enterotoxin and E. coli isolated, 19 (13%) produced enterotoxin respectively.

Conclusions: The contamination of kunu occurs during processing, packaging and by vendors. Improved personal hygiene of the producers, environment and proper preservation methods will reduces microbial proliferation and spoilage of kunu. The consumption of kunu is of public health interest.

Keywords: Bacteria, Contamination, Kunu, Enterotoxin, Strains

1. INTRODUCTION

Kunu or kununzaki is a beverage drink made from grains such as millet, sorghum and maize or their combinations. It is a popular drink in northern parts of Nigeria. Kunu made from sorghum is milky light-brown in colour, while that made from maize is whitish in colour [1,2]. The grain seeds used for the production of kunu drink were allowed to germinate while steeped in water for few days and after which blended with sweet potatoes and ginger or pepper to form a smooth paste. The paste is divided into two, one part is placed in a vessel and boiled water is added to it to form a thick mixture. The unheated half is added to the previous and stirred to give a thick mixture. The mixture is left for a day or two for the grain husk to settle. The husk and other sediments are filtered out of the mixture and the filtrate is boiled for consumption.

Kunu is a non-alcoholic beverage marketed in several public places such as offices, markets, schools, motor parks and a very common consumed beverage in occasions such as weddings, naming ceremonies, birthday celebrations, burials etc.[3] Kunu is an appetizer,
food complement and refresher to quench thirst [4,5,6] The proximate analysis of kunu was determined and the content includes; protein 2.31 – 3.63%, fats 3.35 – 3.65%, ash content 1.16 – 1.21% and carbohydrates 82.92 – 83.55% [2]

There are varieties of Kunu depending on the feed stock used for processing, they are: kunuzaki, kunugyada, kunusamiya, kunubaule, kununjiko and kunugayamba [1,7] Out of these, kununzaki is most widely produced and consumed [8,9] Some of the microorganisms involved in the fermentation of kunu were Lactobacilli, Lactococcus, Enterococcus, Streptococcus, Penicillium and Sacharomyces sp.[10] The high bacterial content of kunu may be an indicator of poor hygiene, poor quality cereals and water used in preparation and packaging processes [11] The bacteria isolated from kunu were E. coli 33.3%, S. aureus 26.7%, Streptococcus sp. 23.3%, Pseudomonas sp. 10% and Bacillus sp. 6.7% [11]. In a study to determine the microbiological quality of kunu in Yenagoa, Bayelsa State, the bacteria isolated were E. coli, Enterobacter sp, Bacillus sp, Salmonella sp, Micrococcius and Streptococcus sp. It was noted that most of the bacteria isolated were of public health importance and they were introduced during processing and handling due to poor hygiene [12] In another research investigating microbial quality of locally produced kunu in Calabar the bacteria isolated were Bacillus sp. 15%, E. coli 15%, Salmonella sp. 12.5%, Streptococcus sp.10%, Pseudomonas sp. 7.5%, Proteus sp. 7.5%, Lacobacillus 22.5% and S. aureus 10% respectively [13]. The microbiological qualities of kunu sold in Calabar were below acceptable standard and unfit for human consumption soughtIt was noted that most of the bacteria isolated were of public health importance and they were introduced during processing and handling due to poor hygiene [14]. Bacteria isolated from kunu might be associated with food spoilage, food infections and poisons [15]. Similar organisms were isolated by other researchers and they attributed the contamination of kunu to processing and handling. The processing and handling of kunu should be improved for consumers wellbeing [16]. Kunuzaki contamination with pathogenic bacteria is of public health importance and might cause diverse food related illnesses and infection to consumers [17]. The aim of this work is to determine the bacteriological quality of kunu sold in Bayelsa, identify isolated bacteria and the enterotoxin producing strains of some isolates.

2. MATERIAL AND METHODS

2.1 Study Area

The study was conducted in Bayelsa State, Nigeria. The samples of Zobo drink were purchased from the three (3) senatorial district headquarters of Bayelsa, namely: Yenagoa (the capital), Sagbama and Ogbia town. Bayelsa state was carved out of River State in 1996. Bayelsa is located in latitude 4°[15] North, latitude 5°23’ South and longitude 5°22’ west and longitude 6°45’ east. It is bound by Delta State on the North, River State on the East and Atlantic Ocean on the West and South. Bayelsa has the largest wetland in West African sub-region. It has a population of about 1.7 million people.

2.2 Collection of Samples

The organism used for the experiment is P. aeruginosa. It was molecularly identified at Lahor Research Laboratories, Benin, Edo State, Nigeria. The organism was stored in 50% glycerol and kept at -20°C.

2.3 Bacteriological Examination of Samples

Each sample of kunu was gently mixed by inversion several times and 1mL of the sample (neat) was added to 9ml of sterile peptone water (sterilized by autoclaving at 121° C for 15 minutes). Subsequent serial dilutions were made up to 105 and 0.0mL of the last dilution
(105) was dropped on already prepared and dried agar plates in duplicates (nutrient, MacC onkey and salmonella/shigella). These were spread evenly on agar media with aid of sterile glass rod (sterilized by dipping in absolute alcohol and flaming in bunsen flame).

The inoculated plates were allowed to dry and incubated at 37° C for 18 - 24 hours before examining for growth.

2.4 Test for Bacterial Load in stored Kunu

A set of freshly prepared kunu were kept at room temperature and another in the refrigerator at about 4° C after the initial determination of the bacteria counts in CFU/mL. The counts from the preserved kunu at room and refrigeration temperature were determined on the second and third day respectively.

2.5 Identification of Isolated Bacteria

The bacteria isolated were identified using morphology, cultural, Gram’s stain reaction, chemical and biochemical reactions such as citrate, VP, Methyl red, indole, catalase, coagulase and carbohydrate fermentation etc.

2.6 Detection of Enterotoxin Producing E. coli from Kunu

PROTM 0157 KIT detects enterotoxin producing E. coli. The Hardy Diagnostics PROTM 0157 KIT provides a rapid latex agglutination method to detect E. coli serogroup 0157 antigen from colonies isolated in the laboratory. These were E. coli producing verotoxin (VT-producing pathogen). Hardy diagnostic E. coli PROTM 0157 Kit contains due latex particles coated with an antiserum against E. coli 0157 antigen. When the coated latex particle is mixed with fresh colonies of E. coli serotype 0157, the bacteria will bind to the antiserum, causing the latex particles to visibly agglutinate, which is indicative of positive result.

2.6.1 Procedure

The reagents were allowed to attain room temperature for about 20 minutes prior to use. Then a drop of sterile saline (Cat. no.K59) was dropped in the circle on the test card and overnight cultures of E. coli were emulsified by mixing it with the saline on the test card. The Latex Reagents were mixed by inverting the tubes several times, prior to use. One (1) drop of E. coli PRO™ O157 Latex Reagent were dispensed onto the test circle on the test card. The Latex Reagent and the organism suspension were then mixed with the wooden applicator provided, using the complete area of the circle. A new stick was used for each reagent. Then the entire card was gently hand-rocked, allowing the mixture to flow slowly over the ring area for up to 2 minutes. Under normal lighting conditions, agglutination (strong clumping) of the latex particles were examined. All organisms yielding a positive agglutination reaction were retested with the Negative Control Latex Reagent.

2.7 Enterotoxicity Testing For S. aureus

The Prolex™ Staph Latex Kit provides a rapid platform for the identification of Staphylococcal isolates particularly S. aureus that produce enterotoxin. The Prolex™ Staph Latex Kit utilizes blue polystyrene latex particles that have been sensitized with fibrinogen and IgG.

2.7.1 Procedure
The test kit was removed from the refrigerator 20 minutes prior to use and the latex reagents were allowed to attain room temperature. The latex reagent was re-suspended by inverting the dropper bottle several times. This was followed by dispensing 1 drop of Staph Test Latex Reagent into a circle on the test card. A sterile loop was used to transfer two colonies of the test isolate into the circle. The reagent and colonies were mixed and spread to cover the entire area of the circle and this was rocked gently on the card allowing the mixture to flow slowly over the entire test ring area. Agglutination was observed for 20 seconds. Negative Control Latex Reagent was included as quality control. Strong agglutination within 20 seconds with the Staph Test Latex Reagent indicates positive result.

3. RESULTS

3.1

A total of 150 samples of kunu were examined for the presence of bacteria. The result obtained showed that from Yenagoa S. aureus was 50 (27.8%), E. coli 50 (27.8%), Bacillus sp. 50 (27.8%), and Streptococcus sp. 30 (16.7%) respectively. From kunu bought from Sagbama, S. aureus were 50 (27.0%), E. coli 50 (28.6%), Bacillus sp. 50 (27.0%) and Streptococcus sp. 35 (18.9%) respectively. Kunu purchased from Ogbia town had S. aureus 50 (28.6%), E. coli 50 (28.6%), Bacillus sp. 50 (28.6%) and Streptococcus sp. 25 (14.3%) respectively. The overall percentage occurrences of isolated bacteria were S. aureus 150 (27.8%), E. coli 150 (27.8%), Bacillus sp. 150 (27.8%) and Streptococcus sp. 90 (16.9%) respectively shown of Table 1.

Table 1. Percentage Occurrence of Bacteria Isolated from Kunu Drink

<table>
<thead>
<tr>
<th>Location</th>
<th>S. aureus</th>
<th>E. coli</th>
<th>Bacillus sp.</th>
<th>Streptococcus sp.</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Yenagoa</td>
<td>50 (27.8)</td>
<td>50 (27.8)</td>
<td>50 (27.8)</td>
<td>30 (16.7)</td>
<td>185 (33.3)</td>
</tr>
<tr>
<td>Sagbama</td>
<td>50 (27.8)</td>
<td>50 (27.8)</td>
<td>50 (27.8)</td>
<td>35 (27.8)</td>
<td>185 (34.6)</td>
</tr>
<tr>
<td>Ogbia</td>
<td>50 (27.8)</td>
<td>50 (27.8)</td>
<td>50 (27.8)</td>
<td>25 (14.3)</td>
<td>175 (32.4)</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td><strong>150 (27.8)</strong></td>
<td><strong>150 (27.8)</strong></td>
<td><strong>150 (27.8)</strong></td>
<td><strong>90 (16.7)</strong></td>
<td><strong>540</strong></td>
</tr>
</tbody>
</table>

Comment [WU4]: Table 1 A legend is needed to indicate the table abbreviations (and information presented in the brackets).
4.2 Enterotoxin Producing S. aureus and E. coli from Kunu

A total of 50 S. aureus were isolated from kunu in Yenegoa out of which 7 (14%) were enterotoxin producing strains and 50 S. aureus were isolated from Sagbama. 10 (20%) produced enterotoxin, while in Ogbia, Town 50 S. aureus were isolated, 8 (16%) were positive for enterotoxin production respectively. Overall total of S. aureus that produced enterotoxin were 25 (16.6%) as shown in Table 2.

Table 2. Percentage Occurrence of Enterotoxin Producing S. aureus and E. coli from Kunu

<table>
<thead>
<tr>
<th>Location</th>
<th>S. aureus</th>
<th>Number positive</th>
<th>E. coli</th>
<th>Number positive</th>
</tr>
</thead>
<tbody>
<tr>
<td>Yenegoa</td>
<td>50</td>
<td>7 (14)</td>
<td>50</td>
<td>7 (14)</td>
</tr>
<tr>
<td>Sagbama</td>
<td>50</td>
<td>10 (20)</td>
<td>50</td>
<td>5 (10)</td>
</tr>
<tr>
<td>Ogbia</td>
<td>50</td>
<td>8 (16)</td>
<td>50</td>
<td>7 (14)</td>
</tr>
<tr>
<td>Total</td>
<td>150</td>
<td>25 (16.6)</td>
<td>150</td>
<td>19 (13)</td>
</tr>
</tbody>
</table>

Numbers in parentheses = percentages

Table 3. Bacterial Counts from Preserved Refrigerated and Non-Refrigerated Kunu (CFU/mL)

<table>
<thead>
<tr>
<th>Temperature</th>
<th>0 Hr</th>
<th>24 Hrs</th>
<th>48 Hrs</th>
</tr>
</thead>
<tbody>
<tr>
<td>Refrigeration</td>
<td>$7.48 \times 10^5$</td>
<td>$8.40 \times 10^5$</td>
<td>$8.72 \times 10^5$</td>
</tr>
<tr>
<td>Room</td>
<td>$7.48 \times 10^5$</td>
<td>$11.96 \times 10^5$</td>
<td>$13 \times 10^5$</td>
</tr>
</tbody>
</table>

P = 0.8298

Fig. Comparison of bacterial counts from 0 Hr, 24Hrs and 48Hrs.
<table>
<thead>
<tr>
<th>S/No</th>
<th>Colour</th>
<th>Surface</th>
<th>Edge</th>
<th>Translucency</th>
<th>Gram Rxn</th>
<th>Size</th>
<th>Shape</th>
<th>Motility</th>
<th>Methyl Red</th>
<th>Voges Proskauer</th>
<th>Oxidase</th>
<th>H2S Production</th>
<th>Indole</th>
<th>Coagulase</th>
<th>Catalase</th>
<th>Citrate</th>
<th>Unase</th>
<th>Starch Hydrolysis</th>
<th>Lactose</th>
<th>Succrose</th>
<th>Maltose</th>
<th>Galactose</th>
<th>Mannitol</th>
<th>Arabinose</th>
<th>Oxidative</th>
<th>Fermentative</th>
<th>Bacilli</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>M</td>
<td>R</td>
<td>E</td>
<td>C</td>
<td>D</td>
<td>+</td>
<td>M</td>
<td>d</td>
<td>R</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>-</td>
<td>Bacillus sp.</td>
</tr>
<tr>
<td>2</td>
<td>C</td>
<td>r</td>
<td>S</td>
<td>E</td>
<td>C</td>
<td>Ml</td>
<td>-</td>
<td>M</td>
<td>d</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>*</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>G</td>
<td>G</td>
<td>G</td>
<td>G</td>
<td>G</td>
<td>G</td>
<td>G</td>
<td>G</td>
<td>-</td>
<td>E coli</td>
</tr>
<tr>
<td>3</td>
<td>C</td>
<td>r</td>
<td>S</td>
<td>E</td>
<td>C</td>
<td>Mt</td>
<td>-</td>
<td>M</td>
<td>d</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>+</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>-</td>
<td>Streptococcus sp.</td>
</tr>
<tr>
<td>4</td>
<td>C</td>
<td>r</td>
<td>S</td>
<td>E</td>
<td>C</td>
<td>Mt</td>
<td>+</td>
<td>M</td>
<td>d</td>
<td>C</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>N</td>
<td>N</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>A</td>
<td>-</td>
<td>S. aureus</td>
</tr>
</tbody>
</table>

187 188

4. DISCUSSION

Comment [WU7]: Table from the lines 188-189 has no number and no matching in text.
The bacteria isolated from kunu drinks in this study were *Staphylococcus* sp, *E. coli*, *Bacillus* sp, and *Streptococcus* sp. The contamination after the boiling process (post CCP contamination) might be responsible for the presence bacteria in kunu. Similar bacteria were isolated from kunu by other researchers in other parts of Nigeria such as *Bacillus* sp., *Salmonella* sp, *Micrococcus*, *Staphylococcus aureus* and *Streptococcus* sp.[13], *E. coli*, *S. aureus*, *Salmonella* sp and *Shigella* sp in Maiduguri [18]. In Kaduna, *Lactobacillus*, *Bacillus* sp., *E. coli*, *S. aureus*, *Salmonella* sp, *Streptococcus* sp.[13].

Other researchers had *S. aureus* 4(10%), *Lactobacillus* 9(22.5%), *Proteus* sp. 3(7.5%), *Streptococcus* sp. 4(10%), *Pseudomonas* sp. 3(7.5%), *Bacillus* sp. 6(15%) and *E. coli* 6(15%) respectively [13]. Most researchers had comparable bacteria in Port Harcourt [7], Oyo and Lagos [1,2], Kano [21], Maiduguri [18], Jalingo [3] in Calabar cross River State [5]. The poor microbial quality of kunu produced locally were because of poor hygiene and poor environmental conditions under which kunu were produced. Health hazard may be associated with consumer of locally produced kunu.

Among the bacteria isolated, Staphylococci were the most prevalent organism. *Staphylococcus* is normal inhabitant of the human body which can be found on the skin, mouth, nostril, hands, various surfaces etc. these were possible sources from where Staphylococci sp. can contaminate kunu during processing and packaging. Bacteria might be present in storage containers, sieve used to filter the finished product and contamination from handlers. The percentage of *S. aureus* was 21.7% [11], while in this study *S. aureus* were 26.7%. The prevalence of *E. coli* was 27.8% from this study but others *E. coli* as 33.3% [11] and *E. coli* as 5.0% [5]. *E. coli* is the most prevalent aerobic bacteria in human and mammal faeces. Contamination by *E. coli* might be by faecal contamination, contaminated water, handlers, processing and packing. *Bacillus* sp. were 27.8% occurrence. *Bacillus* sp. are geophilic and the spores are found in the soil, dust etc. The contamination of grains and spices by *Bacillus* sp. and their spores from soil and dust are likely. The percentage of *Bacillus* sp. isolated other workers were 23.3% [5], and 7.6% [12]. Spores may survive during boiling at about 100°C (the only Critical Control Point (CCP) in the processes of kunu production) and germinate to re-contaminate kunu. Kunu should be preserved under refrigeration temperature at 4°C and or pasteurized to reduce the microbial load and to increase the shelf life [21,5].

It was noted that local drinks such as kunu may act as vehicle for the transmission of zoonotic and bacterial infections such as staphylococcosis, salmonellosis, shigellosis, tuberculosis, listeriosis etc. (Umaru et al., 2014)[3]. Kunu as a beverage sold in public places such as markets, schools, offices etc. is patronized because of the cheap price compared to other soft drinks and is served in occasions such as weddings, naming ceremonies, birthday celebrations for economic reasons and public acceptability. The consumption of contaminated kunu drink may result in outbreak of food borne illness. Preparation of kunu in environment with poor sanitary conditions predisposes the preparation and packaging processes to contamination and exposes the consumers to health risk.

The isolation of 13% of *E. coli* and 17% of *S. aureus* capable of producing enterotoxin indicates that the consumption of kunu is of public health concern. The production of enterotoxin in bacteria is mostly associated with gastrointestinal disturbances and/ or food borne illnesses.

**Conclusions**: The production kunu as non alcoholic beverage for public consumption should be regulated by appropriate regulatory agencies to reduce the risk of consumer infection. Producers should be made to have fair knowledge of food preservation and food sanitation. The isolation of 13% of *E. coli* and 17% of *S. aureus* capable of producing...
enterotoxin indicates that the consumption of locally produced kunu is a public health concern.

REFERENCES


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DEFINITIONS, ACRONYMS, ABBREVIATIONS

Here is the Definitions section. This is an optional section.

Term: Definition for the term